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TITLE: Materials and Methods for Analysis of ATP-binding Cassette Transporter Gene Expression

FIELD OF THE INVENTION

5 The invention relates to materials and methods for detection of ATP-binding cassette transporter gene expression. In particular, the invention relates to primers and the resulting PCR products for detection of ABC transporter gene expression, and the use of said materials and methods in assays and kits.

BACKGROUND OF THE INVENTION

10 ATP-binding cassette (ABC) transporters are one of the largest protein classes known to be involved in the trafficking of biological molecules across membranes. There are 48 different genes in humans which code for ABC transporters. The ABC transporters are classified into families based on the sequence and organization of their ATP-binding domain. Currently, there are seven 15 families, which are designated A through G. The families are further classified into subfamilies based on their gene and protein structure.

20 All of the 48 human genes encoding the ABC transporters have been cloned and sequenced (www.ncbi.nlm.nih.gov; www.humanabc.org). Of these genes, 16 have known function and at least 14 have been associated with a defined human disease.

25 The functional ABC transporters typically contain two nucleotide-binding folds (NBF) and two transmembrane-spanning α -helices. ABC transporters bind to ATP and use the energy from the ATP hydrolysis to drive the transport of various molecules across cell membranes. These transporters are able to transport a variety of compounds across cell membranes against steep concentration gradients. The ABC transporters are involved in the transport of ions, amino acids, peptides, sugars, vitamins, steroid hormones, lipids, bile salts and toxic compounds across cell 30 membranes.

The ABC transporters have been shown to be involved in transporting drugs out of cells, especially anti-cancer drugs. For example ABC B1 (MDR1), ABC C1 (MRP1), ABC C2 (MRP2), and ABC G2 (BCRP) have been characterized and tested for drug resistance. Genetic variations in the ABC transporters may modulate the

phenotype in patients, and thus affect their predisposition to drug toxicity and response to drug treatment (Sparreboom et al., 2003).

The presence of functional ABC transporters in cells may significantly influence the efficacy of drugs. Thus, ABC transporter gene expression experiments 5 in specific cells can be used to tailor drug treatment protocols to specific cell types, tissues, diseases or cancers. For example, a biopsy of a tumor can be tested for the presence of specific ABC transporter gene expression, and the information can be used to choose the most effective drugs for the treatment of that cancer. In addition, the information on ABC transporter gene expression can be used in candidate 10 population profiling, such as the pre-screening of patients for inclusion or exclusion from clinical trials.

There is a need for screening of ABC transporter gene expression, which can be used, for example in drug screening analysis.

SUMMARY OF THE INVENTION

15 The present inventors have prepared primers pairs for the human ABC transporter genes. These primers were used to generate a nucleic acid molecule for the ABC transporter genes, said nucleic acid molecule comprising a sequence that specifically hybridizes to only one of the ABC transporter genes. These nucleic acid molecules have been used in assays to screen for ABC transporter gene expression 20 in test samples.

Accordingly, the present invention includes one or more isolated and purified nucleic acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically hybridizes to one ABC transporter gene. In an embodiment of the invention the one or more nucleic acid molecules comprise a 25 portion of the 3' untranslated region of a human ABC transporter gene. In a further embodiment of the present invention, there is provided a set of at least two nucleic acid molecules, at least 10 nucleic acid molecules, at least 20 nucleic acid molecules, at least 30 nucleic acid molecules or at least 48 nucleic acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically 30 hybridizes to one ABC transporter gene. In another embodiment of the present invention, the set of at least two nucleic acid molecules are attached to a substrate. The substrate may be, for example, a membrane, a glass support, a filter, a tissue culture dish, a polymeric material, a bead or a silica support.

In an embodiment of the present invention, the one or more nucleic acid molecules comprise an isolated and purified nucleic acid sequence selected from those shown in Figures 1 to 47 and Sequence ID NOS: 1 to 47. In a further embodiment of the invention, the one or more nucleic acid molecules comprise an isolated and purified nucleic acid sequence selected from:

- (a) the nucleic acid sequences as shown in SEQ ID NOS: 1 to 47 and Figures 1 to 47, wherein T can also be U;
- (b) nucleic acid sequences complementary to (a);
- (c) nucleic acid sequences which are homologous to (a) or (b); or
- 10 (d) a fragment of (a) to (c), which comprises a sequence that specifically hybridizes to one of the ABC transporter genes.

In an embodiment of the present invention the one or more nucleic acid molecules are prepared from one or more primer pairs using any known amplification method, for example the polymerase chain reaction (PCR). Accordingly, the present 15 invention includes one or more pairs of primers for preparing one or more nucleic acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically hybridizes to one ABC transporter gene. In an embodiment of the present invention, the one or more pairs of primers used to generate such nucleic acid molecules comprise a nucleic acid sequence selected from those listed in Table 20 1 or SEQ ID NOS: 48 to 141. In further embodiments of the invention, the primers comprise:

- (a) the nucleic acid sequences as shown in SEQ ID NOS: 48 to 141 and Table 1, wherein T can also be U;
- (b) nucleic acid sequences complementary to (a); or
- 25 (c) nucleic acid sequences which are homologous to (a) or (b).

In another embodiment of the invention, the primers comprise at least the 5 nucleotides at the 3' end of the sequences as shown in Table 1 or SEQ ID NOS: 48 to 141.

In still further embodiments of the invention, the one or more primers pairs 30 comprise a nucleic acid sequence selected from one or more of:

- (a) SEQ ID NO: 48 and SEQ ID NO: 49;
- SEQ ID NO: 50 and SEQ ID NO: 51;
- SEQ ID NO: 52 and SEQ ID NO: 53;

SEQ ID NO: 54 and SEQ ID NO: 55;
SEQ ID NO: 56 and SEQ ID NO: 57;
SEQ ID NO: 58 and SEQ ID NO: 59;
SEQ ID NO: 60 and SEQ ID NO: 61;
5 SEQ ID NO: 62 and SEQ ID NO: 63;
SEQ ID NO: 64 and SEQ ID NO: 65;
SEQ ID NO: 66 and SEQ ID NO: 67;
SEQ ID NO: 68 and SEQ ID NO: 69;
SEQ ID NO: 70 and SEQ ID NO: 71;
10 SEQ ID NO: 72 and SEQ ID NO: 73;
SEQ ID NO: 74 and SEQ ID NO: 75;
SEQ ID NO: 76 and SEQ ID NO: 77;
SEQ ID NO: 78 and SEQ ID NO: 79;
SEQ ID NO: 80 and SEQ ID NO: 81;
15 SEQ ID NO: 82 and SEQ ID NO: 83;
SEQ ID NO: 84 and SEQ ID NO: 85;
SEQ ID NO: 86 and SEQ ID NO: 87;
SEQ ID NO: 88 and SEQ ID NO: 89;
SEQ ID NO: 90 and SEQ ID NO: 91;
20 SEQ ID NO: 92 and SEQ ID NO: 93;
SEQ ID NO: 94 and SEQ ID NO: 95;
SEQ ID NO: 96 and SEQ ID NO: 97;
SEQ ID NO: 98 and SEQ ID NO: 99;
SEQ ID NO: 100 and SEQ ID NO: 101;
25 SEQ ID NO: 102 and SEQ ID NO: 103;
SEQ ID NO: 104 and SEQ ID NO: 105;
SEQ ID NO: 106 and SEQ ID NO: 107;
SEQ ID NO: 108 and SEQ ID NO: 109;
SEQ ID NO: 110 and SEQ ID NO: 111;
30 SEQ ID NO: 112 and SEQ ID NO: 113;
SEQ ID NO: 114 and SEQ ID NO: 115;
SEQ ID NO: 116 and SEQ ID NO: 117;
SEQ ID NO: 118 and SEQ ID NO: 119;

SEQ ID NO: 120 and SEQ ID NO: 121;
SEQ ID NO: 122 and SEQ ID NO: 123;
SEQ ID NO: 124 and SEQ ID NO: 125;
SEQ ID NO: 126 and SEQ ID NO: 127;
5 SEQ ID NO: 128 and SEQ ID NO: 129;
SEQ ID NO: 130 and SEQ ID NO: 131;
SEQ ID NO: 132 and SEQ ID NO: 133;
SEQ ID NO: 134 and SEQ ID NO: 135;
SEQ ID NO: 136 and SEQ ID NO: 137;
10 SEQ ID NO: 138 and SEQ ID NO: 139; and
SEQ ID NO: 140 and SEQ ID NO: 141;
(b) the nucleic acid sequences in (a) wherein T can also be U;
(c) nucleic acid sequences complementary to (a) or (b); and
(d) nucleic acid sequences which are homologous to (a), (b) or (c).
15 The present invention also includes nucleic acid molecules prepared using PCR and one or more of the pairs of primers of the invention.
Additionally, the invention provides methods for detecting ABC transporter gene expression in general. Accordingly, the present invention includes a method of detecting the expression of one or more ABC transporter genes comprising:
20 (a) providing one or more nucleic acid molecules, each comprising a sequence that specifically hybridizes to one ABC transporter gene;
(b) providing a transcription indicator from a test sample;
(c) allowing the transcription indicator to hybridize with said one or more nucleic acid molecules; and
25 (d) detecting an amount of hybridization of said transcription indicator with said one or more nucleic acid sequences,
wherein the amount of hybridization is indicative of the expression of one or more ABC transporter genes.

In another embodiment of the invention, an array, in particular a microarray is 30 used to detect ABC transporter gene expression in a test sample. Therefore, the present invention also includes an array, in particular a microarray, comprising a substrate and one or more nucleic acid molecules, each comprising a sequence that specifically hybridizes to one ABC transporter gene, wherein said one or more

nucleic acid molecules are immobilized to said substrate. Additionally, the invention provides a method of detecting ABC transporter gene expression in a test sample using a DNA microarray.

The nucleic acid molecules and methods of the present invention can be used 5 to perform drug-associated ABC transporter gene expression profiling.. Such profiling will identify potential modulators of ABC transporter gene expression. Accordingly, in yet another embodiment of the invention, there is provided a method for screening compounds for their effect on the expression of one or more ABC transporter genes comprising:

- 10 (a) exposing a test sample to one or more compounds;
- (b) providing a transcription indicator from the test sample;
- (c) providing one or more nucleic acid sequences, each comprising a sequence that specifically hybridizes to one ABC transporter gene;
- (d) allowing said transcription indicator to hybridize with said one or more 15 nucleic acid sequences; and
- (e) detecting an amount of hybridization of said transcription indicator with said one or more nucleic acid sequences,

wherein the amount of hybridization is indicative of the expression of the one or more ABC transporter genes.

0 In further embodiments, the methods of the invention further comprise (a) generating a set of expression data from the detection of the amount of hybridization; (b) storing the data in a database; and (c) performing comparative analysis on the set of expression data, thereby analyzing ABC transporter gene expression. The present invention also relates to a computer system comprising (a) a database 5 containing information identifying the expression level of a set of genes comprising at least two ABC transporter genes; and (b) a user interface to view the information.

The method for screening compounds for their effect on ABC transporter gene expression is useful for the design of a drugs or chemical therapy for the treatment of disease. In an embodiment, the hybridization assay is a DNA microarray.

0 Other aspects of the present invention include kits for performing the methods of the invention as well as methods of conducting a target discovery business using the methods of the invention.

Other features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples while indicating embodiments of the invention are given by way of illustration only, since various changes and 5 modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will now be described in relation to the drawings in which:

Figure 1 shows a nucleic acid sequence that specifically hybridizes to ABCA1 and

10 corresponds to SEQ ID NO: 1.

Figure 2 shows a nucleic acid sequence that specifically hybridizes to ABCA2 and corresponds to SEQ ID NO: 2.

Figure 3 shows a nucleic acid sequence that specifically hybridizes to ABCA3 and corresponds to SEQ ID NO: 3.

15 Figure 4 shows a nucleic acid sequence that specifically hybridizes to ABCA4 and corresponds to SEQ ID NO: 4.

Figure 5 shows a nucleic acid sequence that specifically hybridizes to ABCA5 and corresponds to SEQ ID NO: 5.

20 Figure 6 shows a nucleic acid sequence that specifically hybridizes to ABCA6 and corresponds to SEQ ID NO: 6.

Figure 7 shows a nucleic acid sequence that specifically hybridizes to ABCA7 and corresponds to SEQ ID NO: 7.

Figure 8 shows a nucleic acid sequence that specifically hybridizes to ABCA8 and corresponds to SEQ ID NO: 8.

25 Figure 9 shows a nucleic acid sequence that specifically hybridizes to ABCA9 and corresponds to SEQ ID NO: 9.

Figure 10 shows a nucleic acid sequence that specifically hybridizes to ABCA10 and corresponds to SEQ ID NO: 10.

30 Figure 11 shows a nucleic acid sequence that specifically hybridizes to ABCA12 and corresponds to SEQ ID NO: 11.

Figure 12 shows a nucleic acid sequence that specifically hybridizes to ABCB1 and corresponds to SEQ ID NO: 12.

Figure 13 shows a nucleic acid sequence that specifically hybridizes to ABCB2 and corresponds to SEQ ID NO: 13.

Figure 14 shows a nucleic acid sequence that specifically hybridizes to ABCB3 and corresponds to SEQ ID NO: 14.

5 Figure 15 shows a nucleic acid sequence that specifically hybridizes to ABCB4 and corresponds to SEQ ID NO: 15.

Figure 16 shows a nucleic acid sequence that specifically hybridizes to ABCB6 and corresponds to SEQ ID NO: 16.

10 Figure 17 shows a nucleic acid sequence that specifically hybridizes to ABCB7 and corresponds to SEQ ID NO: 17.

Figure 18 shows a nucleic acid sequence that specifically hybridizes to ABCB8 and corresponds to SEQ ID NO: 18.

Figure 19 shows a nucleic acid sequence that specifically hybridizes to ABCB9 and corresponds to SEQ ID NO: 19.

15 Figure 20 shows a nucleic acid sequence that specifically hybridizes to ABCB10 and corresponds to SEQ ID NO: 20.

Figure 21 shows a nucleic acid sequence that specifically hybridizes to ABCB11 and corresponds to SEQ ID NO: 21.

20 Figure 22 shows a nucleic acid sequence that specifically hybridizes to ABCC1 and corresponds to SEQ ID NO: 22.

Figure 23 shows a nucleic acid sequence that specifically hybridizes to ABCC2 and corresponds to SEQ ID NO: 23.

Figure 24 shows a nucleic acid sequence that specifically hybridizes to ABCC3 and corresponds to SEQ ID NO: 24.

25 Figure 25 shows a nucleic acid sequence that specifically hybridizes to ABCC4 and corresponds to SEQ ID NO: 25.

Figure 26 shows a nucleic acid sequence that specifically hybridizes to ABCC5 and corresponds to SEQ ID NO: 26.

30 Figure 27 shows a nucleic acid sequence that specifically hybridizes to ABCC6 and corresponds to SEQ ID NO: 27.

Figure 28 shows a nucleic acid sequence that specifically hybridizes to ABCC7 and corresponds to SEQ ID NO: 28.

Figure 29 shows a nucleic acid sequence that specifically hybridizes to ABCC8 and corresponds to SEQ ID NO: 29.

Figure 30 shows a nucleic acid sequence that specifically hybridizes to ABCC9 and corresponds to SEQ ID NO: 30.

5 Figure 31 shows a nucleic acid sequence that specifically hybridizes to ABCC10b and corresponds to SEQ ID NO: 31.

Figure 32 shows a nucleic acid sequence that specifically hybridizes to ABCC11 and corresponds to SEQ ID NO: 32.

10 Figure 33 shows a nucleic acid sequence that specifically hybridizes to ABCC12a and corresponds to SEQ ID NO: 33.

Figure 34 shows a nucleic acid sequence that specifically hybridizes to ABCC13 and corresponds to SEQ ID NO: 34.

Figure 35 shows a nucleic acid sequence that specifically hybridizes to ABCD1 and corresponds to SEQ ID NO: 35.

15 Figure 36 shows a nucleic acid sequence that specifically hybridizes to ABCD2 and corresponds to SEQ ID NO: 36.

Figure 37 shows a nucleic acid sequence that specifically hybridizes to ABCD3 and corresponds to SEQ ID NO: 37.

20 Figure 38 shows a nucleic acid sequence that specifically hybridizes to ABCD4 and corresponds to SEQ ID NO: 38.

Figure 39 shows a nucleic acid sequence that specifically hybridizes to ABCE1 and corresponds to SEQ ID NO: 39.

Figure 40 shows a nucleic acid sequence that specifically hybridizes to ABCF1 and corresponds to SEQ ID NO: 40.

25 Figure 41 shows a nucleic acid sequence that specifically hybridizes to ABCF2 and corresponds to SEQ ID NO: 41.

Figure 42 shows a nucleic acid sequence that specifically hybridizes to ABCF3 and corresponds to SEQ ID NO: 42.

30 Figure 43 shows a nucleic acid sequence that specifically hybridizes to ABCG1 and corresponds to SEQ ID NO: 43.

Figure 44 shows a nucleic acid sequence that specifically hybridizes to ABCG2 and corresponds to SEQ ID NO: 44.

Figure 45 shows a nucleic acid sequence that specifically hybridizes to ABCG4 and corresponds to SEQ ID NO: 45.

Figure 46 shows a nucleic acid sequence that specifically hybridizes to ABCG5 and corresponds to SEQ ID NO: 46.

5 Figure 47 shows a nucleic acid sequence that specifically hybridizes to ABCG8 and corresponds to SEQ ID NO: 47.

Figure 48 shows the ABC transporter gene RT-PCR amplification products from the CaCo2 cell line.

10 Figure 49 shows the ABC transporter gene RT-PCR amplification products from the HEK293 cell line.

Figure 50 shows the ABC transporter gene RT-PCR amplification products from the HepG2 cell line.

Figure 51 shows a fluorescent intensity cluster plot of relative levels of ABC transporter gene expression in various cell lines normalized to GAPDH.

15 Figure 52 shows a fluorescent intensity cluster plot of relative levels of ABC transporter gene expression in various cell lines normalized to actin.

Figure 53 a fluorescent intensity cluster plot of relative levels of ABC transporter gene expression in various cell lines normalized to SH1.

20 Figure 54 shows the relative levels of ABC B1 to B11 gene expression in the HEK cell line normalized to various constitutively expressed control genes.

Figure 55 shows the relative levels of ABC B1 to B11 gene expression in various cell lines.

25 Figure 56 shows a fluorescent intensity cluster plot of relative levels of ABC transporter gene expression in a cell line treated with doxorubicin at various time intervals.

Figure 57 shows a fluorescent intensity cluster plot of relative levels of ABC transporter gene expression in a cell line treated with vinblastine at various time intervals.

0 Figure 58 shows a matrix plot of the relative levels of ABC transporter gene expression in a cell line [HepG2] treated with either doxorubicin [dox] or vinblastine [vin] at various time intervals.

Figure 59 shows a matrix plot of the relative levels of ABC transporter gene expression in several cell lines [A549, CaCo2, HepG2] treated with either acetaminophen [AP] or acetylsalicylic acid [SA].

Figure 60 shows a matrix plot of the relative levels of ABC transporter gene expression in a cell line [A549] treated with either all-trans retinoic acid [AAT], cis-13 retinoic acid [A13], cis-9 retinoic acid [A9] or phorbol-12-myristate-13-acetate [APM].

Figure 61 shows a matrix plot of the relative levels of ABC transporter gene expression in cell lines HTB81 [A], CRL1740 [C] and CRL2505 [D] treated with either no drug [none], methanol [Me], phenobarbital [PhB], acetylsalicylic acid [ASA] or acetaminophen [AAP].

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides materials and methods for detection of ABC transporter gene expression. In particular, the invention relates to nucleic acid molecules for analyzing ABC transporter gene expression, wherein the nucleic acid molecules comprise a sequence that specifically hybridizes to one ABC transporter gene, and methods and materials for obtaining such nucleic acid molecules. The invention also relates to the use of said materials and methods in assays and kits to detect ABC transporter gene expression.

(I) Abbreviations

The following standard abbreviations for the nucleic acid residues are used throughout the specification: A-adenine; C-cytosine; G-guanine; T-thymine; and U-uracil.

(II) Definitions

The term "nucleic acid molecule", "nucleic acid sequence(s)" or "nucleotide sequence" as used herein refers to an oligonucleotide or polynucleotide, and fragments or portions thereof, and to DNA or RNA of genomic or synthetic origin that may be single- or double-stranded, and represent the sense or antisense strand.

The term "ABC transporter genes" refers to nucleic acid sequences encoding the ABC transporters, for example the human ABC transporter genes. There are currently 48 known human transporters, which have been cloned and sequenced (www.ncbi.nlm.nih.gov; www.humanabc.org). The discovery and confirmation of new ABC transporter genes are ongoing. ABC transporter genes in this application are

intended to include unknown ABC transporter genes, which will be discovered or confirmed in the future.

The term "PCR amplicon" refers to a nucleic acid generated by nucleic acid amplification.

5 The term "ABC transporter gene expression" refers to the transcription of an ABC transporter gene into an RNA product.

"Amplification" is defined as the production of additional copies of a nucleic acid sequence and is generally carried out using polymerase chain reaction technologies well known in the art (Dieffenbach CW and GS Dveksler (1995) PCR

10 Primer, a Laboratory Manual, Cold Spring Harbor Press, Plainview N.Y.). As used herein, the term "polymerase chain reaction" ("PCR") refers to the method of K. B. Mullis U.S. Pat. Nos. 4,683,195 and 4,683,202, hereby incorporated by reference, which describe a method for increasing the concentration of a segment of a target sequence in a mixture of genomic DNA without cloning or purification. The length of
15 the amplified segment of the desired target sequence is determined by the relative positions of two oligonucleotide primers with respect to each other, and therefore, this length is a controllable parameter. By virtue of the repeating aspect of the process, the method is referred to as the "polymerase chain reaction" (hereinafter "PCR"). Because the desired amplified segments of the target sequence become the
20 predominant sequences (in terms of concentration) in the mixture, they are said to be "PCR amplified".

Amplification in PCR requires "PCR reagents" or "PCR materials", which herein are defined as all reagents necessary to carry out amplification except the polymerase, primers and template. PCR reagents normally include nucleic acid
25 precursors (dCTP, dTTP etc.) and buffer.

As used herein, the term "primer" refers to an oligonucleotide, whether occurring naturally as in a purified restriction digest or produced synthetically, that is capable of acting as a point of initiation of synthesis when placed under conditions in which synthesis of a primer extension product that is complementary to a nucleic
30 acid strand is induced, (i.e., in the presence of nucleotides and an inducing agent such as DNA polymerase and at a suitable temperature and pH). The primer can be single stranded for maximum efficiency in amplification, but may alternatively be double stranded. If double stranded, the primer is first treated to separate its strands

before being used to prepare extension products. In one embodiment, the primer is an oligodeoxyribonucleotide. The primer must be sufficiently long to prime the synthesis of extension products in the presence of the inducing agent. The exact lengths of the primers will depend on many factors, including temperature, source of 5 primer and the use of the method.

The term "pair(s) of primers" refers to an upper primer and a lower primer. The primers can be categorized as upper or lower primers, depending upon the relative orientation of the primer versus the polarity of the nucleic acid sequence of interest (e.g., whether the primer binds to the coding strand or a complementary 10 (noncoding) strand of the sequence of interest).

The terms "homolog", "homology" and "homologous" as used herein in reference to nucleotides or nucleic acid sequences refer to a degree of complementarity with other nucleotides or nucleic acid sequences. There may be partial homology or complete homology (i.e., identity). A nucleotide sequence that is 15 partially complementary, i.e., "substantially homologous," to a nucleic acid sequence is one that at least partially inhibits a completely complementary sequence from hybridizing to a target nucleic acid sequence. The inhibition of hybridization of the completely complementary sequence to the target sequence may be examined using a hybridization assay (Southern or Northern blot, solution hybridization and the like) 20 under conditions of low stringency. A substantially homologous sequence or probe will compete for and inhibit the binding (i.e., the hybridization) of a completely homologous sequence to a target sequence under conditions of low stringency. This is not to say that conditions of low stringency are such that non-specific binding is 25 permitted; low stringency conditions require that the binding of two sequences to one another be a specific (i.e., selective) interaction. The absence of non-specific binding may be tested by the use of a second target sequence that lacks even a partial degree of complementarity (e.g., less than about 30% identity); in the absence of non-specific binding the probe will not hybridize to the second non-complementary target.

30 Low stringency conditions comprise conditions equivalent to binding or hybridization at 25°C, in a solution consisting of 500mM sodium phosphate pH 6.0, 1% SDS, 1% BSA, 1mM EDTA when a target of about 50 nucleotides in length is employed.

The art knows well that numerous equivalent conditions may be employed to comprise low stringency conditions; factors such as the length and nature (DNA, RNA, base composition) of the probe and nature of the target (DNA, RNA, base composition, present in solution or immobilized, etc.) and the concentration of the 5 salts and other components (e.g., the presence or absence of formamide, dextran sulfate, polyethylene glycol), as well as components of the hybridization solution may be varied to generate conditions of low stringency hybridization different from, but equivalent to, the above listed conditions. In addition, the art knows conditions that 10 promote hybridization under conditions of high stringency (e.g., increasing the temperature of the hybridization and/or wash steps, the use of formamide in the hybridization solution, etc.).

When used in reference to a double-stranded nucleic acid sequence such as a cDNA or genomic clone, the term "substantially homologous" refers to any probe 15 that can hybridize to either or both strands of the double-stranded nucleic acid sequence under conditions of low stringency as described above.

When used in reference to a single-stranded nucleic acid sequence, the term "substantially homologous" refers to any probe that can hybridize (i.e., it is the complement of the single-stranded nucleic acid sequence) under conditions of low stringency as described above.

20 The term "cDNA" refers to complementary or "copy" DNA. Generally, cDNA is synthesized by a DNA polymerase using any type of RNA molecule as a template. Alternatively, the cDNA can be obtained by direct chemical synthesis.

The term "complementary" refers to nucleic acid sequences capable of base-pairing according to the standard Watson-Crick complementary rules, or being 25 capable of hybridizing to a particular nucleic acid segment under stringent conditions.

The term "hybridization" refers to duplex formation between two or more polynucleotides to form, for example a double-stranded nucleic acid, via base pairing. The ability of two regions of complementarity to hybridize and remain 30 together depends on the length and continuity of the complementary regions, and the stringency of the hybridization conditions.

The term "DNA microarray" refers to substrate with at least one target DNA immobilized to said substrate. The target DNA molecules are typically immobilized in

prearranged patterns so that their locations are known or determinable. Nucleic acids in a sample can be detected by contacting the sample with the DNA microarray; allowing the target DNA and nucleic acids in the sample to hybridize; and analyzing the extent of hybridization.

5 The term "label" refers to any detectable moiety. A label may be used to distinguish a particular nucleic acid from others that are unlabelled, or labeled differently, or the label may be used to enhance detection.

10 The term "nucleic acids" refers to a polymer of ribonucleic acids or deoxyribonucleic acids, including RNA, mRNA, rRNA, tRNA, small nuclear RNAs, cDNA, DNA, PNA, or RNA/DNA copolymers. Nucleic acid may be obtained from a 15 cellular extract, genomic or extragenomic DNA, viral RNA or DNA, or artificially/chemically synthesized molecules.

15 The term "RNA" refers to a polymer of ribonucleic acids, including RNA, mRNA, rRNA, tRNA and small nuclear RNAs, as well as to RNAs that comprise ribonucleotide analogues to natural ribonucleic acid residues, such as 2-O-methylated residues.

20 The term "transcription" refers to the process of copying a DNA sequence of a gene into an RNA product, generally conducted by a DNA-directed RNA polymerase using the DNA as a template.

25 The term "isolated" when used in relation to a nucleic acid molecule or sequence, refers to a nucleic acid sequence that is identified and separated from at least one contaminant nucleic acid with which it is ordinarily associated in its natural source. Isolated nucleic acid is nucleic acid present in a form or setting that is different from that in which it is found in nature.

30 As used herein, the term "purified" or "to purify" refers to the removal of undesired components from a sample.

As used herein, the term "substantially purified" refers to molecules, either nucleic or amino acid sequences, that are removed from their natural environment, isolated or separated, and are at least 60% free, 75% free, or 90% free from other components with which they are naturally associated. An "isolated nucleic acid molecule" is therefore a substantially purified nucleic acid molecule.

(III) Nucleic Acid Molecules

The present invention provides one or more isolated and purified nucleic acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically hybridizes to only one ABC transporter gene. By "specifically hybridizes to" it is meant that the subject nucleic acid sequence will bind, duplex or hybridize substantially to or only with a particular nucleic acid sequence with minimum cross-hybridization with the other members of this gene family. In other words, the nucleic acid sequence represents a probe for one ABC transporter gene. In an embodiment of the invention, the one or more nucleic acid molecules comprise a portion of the 3' untranslated region of a human ABC transporter gene.

In a further embodiment of the present invention, there is provided a set of at least two nucleic acid molecules, at least 10 nucleic acid molecules, at least 20 nucleic acid molecules, at least 30 nucleic acid molecules or at least 48 nucleic acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically hybridizes to one ABC transporter gene. In another embodiment of the present invention, the set of at least two nucleic acid molecules are attached to a substrate. The substrate may be, for example, a membrane, a glass support, a filter, a tissue culture dish, a polymeric material, a bead or a silica support.

In an embodiment of the present invention, the one or more nucleic acid molecules comprise an isolated and purified nucleic acid sequence selected from those shown in Figures 1 to 47 and Sequence ID NOS: 1 to 47. In a further embodiment of the invention, the one or more nucleic acid molecules comprise an isolated and purified nucleic acid sequence selected from:

- (a) the nucleic acid sequences as shown in SEQ ID NOS: 1 to 47 and Figures 1 to 47, wherein T can also be U;
- (b) nucleic acid sequences complementary to (a);
- (c) nucleic acid sequences which are homologous to (a) or (b); or
- (d) a fragment of (a) to (c), which comprises a sequence that specifically hybridizes to one of the ABC transporter genes.

In an embodiment of the present invention the one or more nucleic acid molecules are prepared from one or more primer pairs using any known amplification method, for example the polymerase chain reaction (PCR). Accordingly, the present invention includes one or more pairs of primers for preparing one or more nucleic

acid molecules, wherein each of the nucleic acid molecules comprises a sequence that specifically hybridizes to one ABC transporter gene. In an embodiment of the present invention, the one or more pairs of primers used to generate such nucleic acid molecules comprise a nucleic acid sequence selected from those listed in Table 5 1 or SEQ ID NOS: 49 to 144. In further embodiments of the invention, the primers comprise:

- (a) the nucleic acid sequences as shown in SEQ ID NOS: 48 to 141 and Table 1, wherein T can also be U;
- (b) nucleic acid sequences complementary to (a); or
- 10 (c) nucleic acid sequences which are homologous to (a) or (b).

In another embodiment of the invention, the primers comprise at least the 5 nucleotides at the 3' end of the sequences as shown in Table 1 or SEQ ID NOS: 48 to 141.

In still further embodiments of the invention, the one or more primers pairs 15 comprise a nucleic acid sequence selected from one or more of:

- (a) one or more isolated and purified pairs of nucleic acid sequences selected from:

SEQ ID NO: 48 and SEQ ID NO: 49;

SEQ ID NO: 50 and SEQ ID NO: 51;

20 SEQ ID NO: 52 and SEQ ID NO: 53;

SEQ ID NO: 54 and SEQ ID NO: 55;

SEQ ID NO: 56 and SEQ ID NO: 57;

SEQ ID NO: 58 and SEQ ID NO: 59;

SEQ ID NO: 60 and SEQ ID NO: 61;

25 SEQ ID NO: 62 and SEQ ID NO: 63;

SEQ ID NO: 64 and SEQ ID NO: 65;

SEQ ID NO: 66 and SEQ ID NO: 67;

SEQ ID NO: 68 and SEQ ID NO: 69;

SEQ ID NO: 70 and SEQ ID NO: 71;

30 SEQ ID NO: 72 and SEQ ID NO: 73;

SEQ ID NO: 74 and SEQ ID NO: 75;

SEQ ID NO: 76 and SEQ ID NO: 77;

SEQ ID NO: 78 and SEQ ID NO: 79;

SEQ ID NO: 80 and SEQ ID NO: 81;
SEQ ID NO: 82 and SEQ ID NO: 83;
SEQ ID NO: 84 and SEQ ID NO: 85;
SEQ ID NO: 86 and SEQ ID NO: 87;
5 SEQ ID NO: 88 and SEQ ID NO: 89;
SEQ ID NO: 90 and SEQ ID NO: 91;
SEQ ID NO: 92 and SEQ ID NO: 93;
SEQ ID NO: 94 and SEQ ID NO: 95;
SEQ ID NO: 96 and SEQ ID NO: 97;
10 SEQ ID NO: 98 and SEQ ID NO: 99;
SEQ ID NO: 100 and SEQ ID NO: 101;
SEQ ID NO: 102 and SEQ ID NO: 103;
SEQ ID NO: 104 and SEQ ID NO: 105;
SEQ ID NO: 106 and SEQ ID NO: 107;
15 SEQ ID NO: 108 and SEQ ID NO: 109;
SEQ ID NO: 110 and SEQ ID NO: 111;
SEQ ID NO: 112 and SEQ ID NO: 113;
SEQ ID NO: 114 and SEQ ID NO: 115;
SEQ ID NO: 116 and SEQ ID NO: 117;
20 SEQ ID NO: 118 and SEQ ID NO: 119;
SEQ ID NO: 120 and SEQ ID NO: 121;
SEQ ID NO: 122 and SEQ ID NO: 123;
SEQ ID NO: 124 and SEQ ID NO: 125;
SEQ ID NO: 126 and SEQ ID NO: 127;
25 SEQ ID NO: 128 and SEQ ID NO: 129;
SEQ ID NO: 130 and SEQ ID NO: 131;
SEQ ID NO: 132 and SEQ ID NO: 133;
SEQ ID NO: 134 and SEQ ID NO: 135;
SEQ ID NO: 136 and SEQ ID NO: 137;
30 SEQ ID NO: 138 and SEQ ID NO: 139; and
SEQ ID NO: 140 and SEQ ID NO: 141;
(b) the nucleic acid sequences in (a) wherein T can also be U;
(c) nucleic acid sequences complementary to (a) or (b); and

(d) nucleic acid sequences which are homologous to (a), (b) or (c).

The present invention also includes nucleic acid molecules prepared using PCR and one or more of the pairs of primers of the invention.

(IV) Method for detecting ABC transporter gene expression

5 Transcription of genes into RNA is a critical step in gene expression. Therefore gene expression can be monitored by monitoring various transcription indicators. There are a variety of techniques known in the art to analyze and quantify gene transcription. In an embodiment of the present invention, ABC transporter gene expression was detected by monitoring or detecting the
10 hybridization of transcription indicators from a test sample with the one or more nucleic acid molecules of the present invention, wherein the one or more nucleic acid molecules comprise a sequence that specifically hybridizes to one ABC transporter gene. In an embodiment, ABC transporter gene expression was detected using reverse transcription. For example, RNA was extracted from a test sample using
15 techniques known in the art. cDNA was then synthesized using known techniques, such as using either oligo(dT) or random primers. ABC transporter gene expression was then detected using the said cDNA by allowing the cDNA to hybridize to the one or more nucleic acid molecules, then detecting the amount of hybridization of said cDNA with the one or more nucleic acid molecules.

20 Accordingly, the present invention includes a method of detecting the expression of one or more ABC transporter genes comprising:

(a) providing one or more nucleic acid molecules, each comprising a sequence that specifically hybridizes to one ABC transporter gene;

(a) providing transcription indicators from a test sample;

25 (b) allowing the transcription indicators to hybridize with said one or more nucleic acid molecules; and

(c) detecting an amount of hybridization of said transcription indicators with said one or more nucleic acid sequences,

wherein the amount of hybridization is indicative of the expression of one or more
30 ABC transporter genes.

(a) Transcription indicators

One of skill in the art will appreciate that it is desirable to have transcription indicators from a test sample that contain suitable nucleic samples having target

nucleic acid sequences that reflect the transcripts of interest. Therefore, suitable nucleic acid samples from the test sample may contain transcripts of interest. Suitable nucleic acid samples, however, may contain nucleic acids derived from the transcripts of interest. As used herein, a nucleic acid derived from a transcript refers 5 to a nucleic acid for whose synthesis the mRNA transcript or a subsequence thereof has ultimately served as a template. Thus, a cDNA reverse transcribed from a transcript, an RNA transcribed from that cDNA, a DNA amplified from the cDNA, an RNA transcribed from the amplified DNA, etc., are all derived from the transcript and detection of such derived products is indicative of the presence and/or abundance of 10 the original transcript in a sample. Thus, suitable transcription indicators include, but are not limited to, transcripts of the gene or genes, cDNA reverse transcribed from the transcript, cRNA transcribed from the cDNA, DNA amplified from the genes, RNA transcribed from amplified DNA, and the like. In an embodiment the transcription indicator is cDNA.

15 Transcripts, as used herein, may include, but not limited to pre-mRNA nascent transcript(s), transcript processing intermediates, mature mRNA(s) and degradation products. It is not necessary to monitor all types of transcripts to practice this invention. For example, one may choose to practice the invention to measure the mature mRNA levels only.

20 The term "test sample" refers to one or more cells, cell lines, tissues or organisms, or fragments thereof. In one embodiment, the test sample is from a human. In an embodiment of the present invention, the test sample is a homogenate of cells or tissues or other biological samples. For example, such sample can be a total RNA preparation of a biological sample or such a nucleic acid sample can be 25 the total mRNA isolated from a biological sample. Those of skill in the art will appreciate that the total mRNA prepared with most methods includes not only the mature mRNA, but also the RNA processing intermediates and nascent pre-mRNA transcripts. For example, total mRNA purified with a poly (dT) column contains RNA molecules with poly (A) tails. Those polyA+ RNA molecules could be mature mRNA, 0 RNA processing intermediates, nascent transcripts or degradation intermediates.

In an embodiment of the present invention, the test sample is a clinical sample with is a sample derived from a patient. Typical clinical samples include, but are not limited to, sputum, blood, blood cells (e.g. white blood cells), tissue or fine

needle biopsy samples, urine, peritoneal fluid and pleural fluid, or cells therefrom. In another embodiment of the present invention, the test sample is derived from a cell culture containing specific cell lines, for example, HepG2, CaCo2 or HEK 293.

One skilled in the art will appreciate that one can inhibit or destroy RNase 5 present in any sample before they are used in the methods of the invention. Methods of inhibiting or destroying nucleases, including RNase, are well known in the art. For example, chaotropic agents may be used to inhibit nucleases or, alternatively, heat treatment followed by proteinase treatment may be used.

Methods of isolating total mRNA are also well known to those skilled in the 10 art. For example, see Chapter 3 of *Laboratory Techniques in Biochemistry and Molecular Biology: Hybridization with Nucleic Acid Probes, Part I: Theory and Nucleic Acid Preparation*, Tijssen, ed. Elsevier Press (1993); Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2nd ed.), Vols. 1-3, Cold Spring Harbour 15 Laboratory (1989); or *Current Protocols in Molecular Biology*, F. Ausubel *et al.*, ed. Greene Publishing and Wiley-Interscience, New York (1987). In an embodiment, the total RNA is isolated from a given test sample, for example, using TRIzol reagent (Cat. No. 15596-018, Invitrogen Life Technologies) according to the manufacturer's instructions.

In embodiments of the present invention, the transcription indicator, whether it 20 be cDNA or mRNA, may need to be amplified prior to performing the hybridization assay. Methods for amplification, including "quantitative amplification" are well known to those skilled in the art.

In an embodiment the transcription indicator is labeled with a detectable label. 25 Methods for labeling nucleic acids are well known to those skilled in the art. In an embodiment of the invention, the label is simultaneously incorporated during an amplification step in the preparation of the transcription indicators. Thus for example, PCR with labeled primers or labeled nucleotides (for example fluorescein-labeled UTP and/or CTP) will provide a labeled amplification product. Alternatively, a 30 label may be added directly to the original nucleic acid sample or to the amplification product after the amplification is completed using methods known to those skilled in the art (for example nick translation and end-labeling).

Detectable labels that are suitable for use in the methods of the present invention, include those that are detectable by spectroscopic, photochemical,

biochemical, immunochemical, electrical, optical or other means. Some examples of useful labels include biotin staining with labeled streptavidin conjugate, magnetic beads, fluorescent dyes (e.g. fluorescein, rhodamine, green fluorescent protein and the like), radiolabels (e.g. ^3H , ^{32}P , ^{14}C , ^{25}S or ^{125}I), enzymes (e.g. horseradish peroxidase, alkaline phosphatase and others commonly used in ELISA) and colorimetric labels such as colloidal gold or colored glass or plastic (e.g. polystyrene, polypropylene, latex and the like) beads. Patents teaching the use of such labels include U.S. Patent Nos. 3,817,837, 3,850,752, 3,939,350, 3,996,345, 4,277,437, 4,275,149 and 4,366,241, the contents of all of which are incorporated herein by reference.

(b) Assay Format

The method of detecting ABC transporter gene expression can be performed using any hybridization assay, including solution and solid phase. Typically a set containing two or more nucleic acid molecules of the invention, each of said nucleic acid molecules comprising a sequence that specifically hybridizes to one ABC transporter gene, are put together in a common container or on a common object. These may be on an array or in a kit together. They are typically separated, either spatially on a solid support such as an array, or in separate vessels, such as vials, tubes or wells in a microwell plate.

According to the present invention, at least 5% of the nucleic acid molecules or probes in a set comprise a sequence that specifically hybridizes to one ABC transporter gene. In an embodiment, more than 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or 95% of such nucleic acid molecules or probes in the set comprise a sequence that specifically hybridizes to one ABC transporter gene.

In an embodiment of the present invention the method of detecting ABC transported gene expression is performed in an array format. One of skill in the art will appreciate that an enormous number of array designs are suitable for the practice of this invention. The array will typically include a number of nucleic acid molecules or probes that specifically hybridize to the sequences of interest. In addition, in an embodiment, the array will include one or more control nucleic acid molecules or probes. The control probes may be, for example, expression level controls (e.g. positive controls and background negative controls).

Background controls are elements printed on the substrate that contain no nucleic acids and thus measure the amount of non-specific hybridization of the labelled cDNA to elements on the substrate.

Expression level controls are probes that hybridize specifically with 5 constitutively expressed genes in the biological sample. Virtually any constitutively expressed gene provides a suitable target for expression level controls. Typically expression level control probes have sequences complementary to subsequences of constitutively expressed "housekeeping genes" including, but not limited to the *beta*-actin gene, the transferrin receptor gene, the glyceraldehyde-3-phosphate 10 dehydrogenase (GAPDH) gene, and the like [Warrington JA *et al.*, *Physiol Genomics* 2:143-147, 2000, Hsiao LL *et al.*, *Physiol Genomics* 7:97-104, 2001, Whitfield ML *et al.*, *Mol Cell Biol* 13:1977-2000, 2002].

In embodiments of the invention the method of detecting ABC transporter expression in a test sample is performed once or more, over a set period of time and 15 at specified intervals, to monitor ABC transporter expression over that period of time.

DNA microarrays have the benefit of assaying gene expression in a high throughput fashion. These microarrays comprise short nucleic acid sequences that are immobilized on or directly chemically synthesized on a substrate, which can then be used in a hybridization reaction with nucleotides extracted from a test sample.

20 Microarrays have the advantage of being able to measure the expression level of hundreds of genes simultaneously.

Accordingly, in an embodiment of the present invention there is provided a DNA microarray comprising one or more nucleic acid molecules arrayed on a substrate, wherein each of the one or more nucleic acid molecules comprise a 25 sequence that specifically hybridizes to one ABC transporter gene. In an embodiment of the invention, the one or more nucleic acid molecules are selected from:

- (a) the nucleic acid sequences as shown in SEQ ID NOS: 1 to 47 and Figures 1 to 47, wherein T can also be U;
- 30 (b) nucleic acid sequences complementary to (a);
- (c) nucleic acid sequences which are homologous to (a) or (b); and
- (d) a fragment of (a) to (c), which comprises a sequence that specifically hybridizes to one of the ABC transporter genes, or

one or more nucleic acids prepared using PCR and one or more primer pairs selected from:

(a) SEQ ID NO: 48 and SEQ ID NO: 49;
SEQ ID NO: 50 and SEQ ID NO: 51;
5 SEQ ID NO: 52 and SEQ ID NO: 53;
SEQ ID NO: 54 and SEQ ID NO: 55;
SEQ ID NO: 56 and SEQ ID NO: 57;
SEQ ID NO: 58 and SEQ ID NO: 59;
SEQ ID NO: 60 and SEQ ID NO: 61;
10 SEQ ID NO: 62 and SEQ ID NO: 63;
SEQ ID NO: 64 and SEQ ID NO: 65;
SEQ ID NO: 66 and SEQ ID NO: 67;
SEQ ID NO: 68 and SEQ ID NO: 69;
SEQ ID NO: 70 and SEQ ID NO: 71;
15 SEQ ID NO: 72 and SEQ ID NO: 73;
SEQ ID NO: 74 and SEQ ID NO: 75;
SEQ ID NO: 76 and SEQ ID NO: 77;
SEQ ID NO: 78 and SEQ ID NO: 79;
SEQ ID NO: 80 and SEQ ID NO: 81;
20 SEQ ID NO: 82 and SEQ ID NO: 83;
SEQ ID NO: 84 and SEQ ID NO: 85;
SEQ ID NO: 86 and SEQ ID NO: 87;
SEQ ID NO: 88 and SEQ ID NO: 89;
SEQ ID NO: 90 and SEQ ID NO: 91;
25 SEQ ID NO: 92 and SEQ ID NO: 93;
SEQ ID NO: 94 and SEQ ID NO: 95;
SEQ ID NO: 96 and SEQ ID NO: 97;
SEQ ID NO: 98 and SEQ ID NO: 99;
SEQ ID NO: 100 and SEQ ID NO: 101;
30 SEQ ID NO: 102 and SEQ ID NO: 103;
SEQ ID NO: 104 and SEQ ID NO: 105;
SEQ ID NO: 106 and SEQ ID NO: 107;
SEQ ID NO: 108 and SEQ ID NO: 109;

SEQ ID NO: 110 and SEQ ID NO: 111;

SEQ ID NO: 112 and SEQ ID NO: 113;

SEQ ID NO: 114 and SEQ ID NO: 115;

SEQ ID NO: 116 and SEQ ID NO: 117;

5 SEQ ID NO: 118 and SEQ ID NO: 119;

SEQ ID NO: 120 and SEQ ID NO: 121;

SEQ ID NO: 122 and SEQ ID NO: 123;

SEQ ID NO: 124 and SEQ ID NO: 125;

SEQ ID NO: 126 and SEQ ID NO: 127;

10 SEQ ID NO: 128 and SEQ ID NO: 129;

SEQ ID NO: 130 and SEQ ID NO: 131;

SEQ ID NO: 132 and SEQ ID NO: 133;

SEQ ID NO: 134 and SEQ ID NO: 135;

SEQ ID NO: 136 and SEQ ID NO: 137;

15 SEQ ID NO: 138 and SEQ ID NO: 139; and

SEQ ID NO: 140 and SEQ ID NO: 141;

(b) the nucleic acid sequences in (a) wherein T can also be U;

(c) nucleic acid sequences complementary to (a) or (b); and

(d) nucleic acid sequences which are homologous to (a), (b) or (c).

20 In embodiments of the invention, the one or more nucleic acid molecules are arranged in distinct spots that are known or determinable locations within the array on the substrate. A spot refers to a region of target DNA attached to the substrate as a result of contacting a solution comprising target DNA with the substrate. Each spot can be sufficiently separated from each other spot on the substrate such that 25 they are distinguishable from each other during the hybridization analysis. In an embodiment, there are at least 48 spots on the DNA microarray; one spot for each of the 48 PCR products generated by the 48 sets of primers disclosed herein which are used as target DNA. In another embodiment, the DNA microarray includes at least one spot for an expression level control as described herein above.

30 The substrate may be any solid support to which nucleic acids can be immobilized, such as a membrane, a glass support, a filter, a tissue culture dish, a polymeric material, a bead or a silica support. For example, the substrate can be a NoAb BioDiscoveries Inc. activated covalent-binding epoxy slide [UAS0005E].

When the nucleic acid molecule is immobilized on the substrate, a conventionally known technique can be used. For example, the surface of the substrate can be treated with polycations such as polylysines to electrostatically bind the target molecules through their charges on the surface of the substrate, and 5 techniques to covalently bind the 5'-end of the target DNA to the substrate may be used. Also, a substrate that has linkers on its surface can be produced, and functional groups that can form covalent bonds with the linkers can be introduced at the end of the DNA to be immobilized. Then, by forming a covalent bond between the linker and the functional group, the DNA and such can be immobilized.

10 Other methods of forming arrays of oligonucleotides, peptides and other polymer sequences with a minimal number of synthetic steps are known and may be used in the present invention. These methods include, but are not limited to, light-directed chemical coupling and mechanically directed coupling. See Pirrung et al., U.S. Patent No. 5,143,854 and PCT Application No. WO 90/15070, Fodor et al., PCT 15 Publication Nos. WO 92/10092 and WO 93/09668, which disclose methods of forming vast arrays of peptides, oligonucleotides and other molecules using, for example, light-directed synthesis techniques. See also, Fodor et al., *Science*, 251, 767-77 (1991). These procedures for synthesis of polymer arrays are now referred 20 to as VLSIPSTM procedures. Using the VLSIPSTM approach, one heterogeneous array of polymers is converted, through simultaneous coupling at a number of reaction sites, into a different heterogeneous array.

Transcription indicators (targets) from a test sample that have been subjected to particular stringency conditions hybridize to the nucleic acid molecules (probes) on the array. One of skill in the art will appreciate that hybridization conditions may be 15 selected to provide any degree of stringency. In an embodiment, hybridization is performed at low stringency [15-18hrs at 37°C in 500mM sodium Phosphate pH 6.0, 1% SDS, 1% BSA, 1mM EDTA] to ensure hybridization and then subsequent washes are performed at higher stringency [0.1xSSC;0.1%SDS then 0.1xSSC then water] to eliminate mismatched hybrid duplexes. Successive washes may be 0 performed at increasingly higher stringency until a desired level of hybridization specificity is obtained. Stringency can also be increased by addition of agents such as formamide. Hybridization specificity may be evaluated by comparison of hybridization to the test nucleic acid sequences with hybridization to the various

controls that can be present (e.g., expression level controls (positive and negative), etc.).

The nucleic acids that do not form hybrid duplexes are washed away leaving the hybridized nucleic acids to be detected, typically through detection of an attached 5 detectable label. After hybridization, the arrays are inserted into a scanner that can detect patterns of hybridization. These patterns are detected by detecting the labeled transcription indicator now attached to the array, for e.g., if the transcription indicator is fluorescently labeled, the hybridization data are collected as light emitted from the labeled groups. Comparison of the absolute intensities of an array hybridized to 10 nucleic acids from a test sample with intensities produced from the various control samples provides a measure of the relative expression of the nucleic acids represented by each of the probes.

If the transcription indicator, for example cDNA, is fluorescently labeled, the fluorescence is detected and acquired using a fluorescence scanner, for example, a 15 GSI Lumonics ScanArray Lite Microarray Analysis System, and the fluorescence intensity analyzed with specific quantitation and data processing software on a dedicated computer, for example, QuantArray and GeneLinker Gold. In an embodiment, the intensity of fluorescence increases with increased ABC transporter gene expression. If the transcription indicator, for example cDNA, is radiolabelled, 20 then detection can be carried out using an RU image scanner and such, and the intensity of the radiation can be analyzed with a computer. In an embodiment, the intensity of the radiation increases with increased ABC transporter gene expression.

In further embodiments of the present invention, the methods of the invention further comprise (a) generating a set of expression data from the detection of the 25 amount of hybridization; (b) storing the data in a database; and (c) performing comparative analysis on the set of expression data, thereby analyzing ABC transporter gene expression. The present invention also relates to a computer system comprising (a) a database containing information identifying the expression level of a set of genes comprising at least two ABC transporter genes; and b) a user 30 interface to view the information.

(V) Drug Screening Assays

In one embodiment, the method of the invention has been used in a drug screening analysis. For example, a test sample was exposed to a chemical

compound or a drug, and then ABC transporter gene expression was detected in the test sample using the methods of the invention. In an embodiment of the invention, ABC transporter expression was detected at various time intervals after the test sample was exposed to a compound or drug, for example every 2 hours after 5 exposure for 24 hours. In a further embodiment, after the test sample was exposed to the chemical or drug, mRNA was extracted from the test sample and then cDNA was produced using the extracted mRNA. The cDNA was labeled and allowed to hybridize with the one or more nucleic acid molecules, wherein each one of the one or more nucleic acid molecules comprised a sequence that specifically hybridizes to 10 one ABC transporter gene. The amount of hybridization was detected and compared with the amount of hybridization obtained with the test sample treated under the same conditions except that it had not been exposed to the compound or drug (i.e. a control sample). By performing this comparison, the effect of the drug or compound on the expression of each of the ABC transporter genes (whether it be 15 increased, decreased or the same) was determined.

Therefore, the nucleic acid molecules and methods of the present invention can be used to perform drug-associated ABC transporter gene expression profiling. Such profiling will identify potential modulators of ABC transporter gene expression. Accordingly, in yet another embodiment of the invention, there is provided a method 20 for screening compounds for their effect on the expression of one or more ABC transporter genes comprising:

- (a) exposing a test sample to one or more compounds;
- (b) providing a transcription indicator from the test sample;
- (c) providing one or more nucleic acid sequences, each comprising a 25 sequence that specifically hybridizes to one ABC transporter gene;
- (d) allowing said transcription inhibitor to hybridize with said one or more nucleic acid sequences; and
- (e) detecting an amount of hybridization of said transcription indicator with said one or more nucleic acid sequences,

30 wherein the amount of hybridization is indicative of expression of the one or more ABC transporter genes.

In further embodiments of the invention the method for screening compounds for their effect on the expression of one or more ABC transporter genes further comprises the steps of

5 (f) quantitatively or qualitatively comparing the amount of hybridization detected in step (e) with the amount of hybridization of transcription indicators from a control sample, thereby determining the effect of the one or more compounds on the expression of the one or more ABC transporter genes.

10 The term "control sample" as used herein means a sample that has been treated under the same conditions as the test sample except that it has not been exposed to one or more compounds, drugs or other conditions that may have an effect on ABC transporter gene expression.

15 The term "compound" as used herein means any agent, including drugs, which may have an effect of ABC transporter gene expression and includes, but is not limited to, small inorganic or organic molecules: peptides and proteins and fragments thereof; carbohydrates, and nucleic acid molecules and fragments thereof. The compound may be isolated from a natural source or be synthetic. The term compound also includes mixtures of compounds or agents such as, but not limited to, combinatorial libraries and extracts from an organism.

20 The term "exposed" as used herein means that the sample has been brought into contact with the compound(s) using any method known in the art. For example, cells lines may be exposed to a compound by adding the compound(s) to the media used for cell storage, growth and/or washing. In a further example, the exposure may be effected by administering the compound(s) to a test subject using any known methods for administration, and the test sample is obtained from the subject, again using any known means.

25 In a further embodiment of the present invention there is provided a method for screening compounds for their effect on the expression of one or more ABC transporter genes comprising:

30 (a) preparing an ABC transporter gene expression profile, using a method of the invention, of a test sample that has been exposed to the compound;
(b) preparing an ABC transporter gene expression profile, using a method of the invention, of a control sample; and

(c) quantitatively or qualitatively comparing the gene expression profile in (a) and (b),

wherein differential expression in (a) and (b) is indicative of a compound having an effect on the expression of one or more ABC transporter genes.

5 In yet another embodiment of the invention, the expression of one or more ABC transporter genes in the test and/or control samples is monitored over a set period of time and at specified time intervals to determine the effect of the compound on the expression of one or more ABC transporter genes over that period of time.

10 In embodiments of the invention, the methods may be used to identify compounds or agents that stimulate, induce and/or up-regulate the transcription or expression of one or more ABC transporter genes, or to down-regulate, suppress and/or counteract the transcription or expression of one or more ABC transporter genes, or that have no effect on transcription or expression of one or more ABC transporter genes, in a given system. According to the present invention, one can 15 also compare the specificity of a compound's effect by looking at the number of ABC transporter genes, the expression of which has been effected. More specific compounds will have fewer transcriptional targets. Further, similar sets of results for two different compounds indicates a similarity of effects for the two compounds.

20 The ABC expression data can be used to design or choose an effective drug or chemical for the treatment of disease, such as cancer. By knowing which of the ABC transporter genes are modulated in the presence of the drug or compound, one can determine a cell's or patient's predisposition to drug toxicity and/or response to drug treatment. For example, if the chemical or drug up-regulates or increases the expression of certain ABC transporters in a test sample that are known to be 25 involved in transporting compounds out of cells, for example ABC B1 (MDR1), ABC C1 (MRP1), ABC C2 (MRP2), or ABC G2 (BCRP), then the efficacy of that compound may be lowered. Further, if the compound down-regulates or decreases the expression of certain ABC transporters in a test sample that are known to be involved in transporting compounds out of cells, for example ABC B1 (MDR1), ABC 30 C1 (MRP1), ABC C2 (MRP2), or ABC G2 (BCRP), then the efficacy and/or toxicity of that compound may be increased.

Accordingly the present invention further relates to a method of assessing the toxicity and/or efficacy of a compound comprising:

(a) preparing an ABC transporter gene expression profile, using a method of the invention, of a test sample that has been exposed to the compound;

(b) preparing an ABC transporter gene expression profile, using a method of the invention, of a control sample; and

5 (c) quantitatively or qualitatively comparing the ABC transporter gene expression profile from (a) and (b),

wherein a difference in the ABC transporter gene expression profiles in (a) and (b) is indicative of the toxicity and/or efficacy of the compound.

In an embodiment of the invention, if the expression of one or more of the 10 ABC transporter genes in the test sample is increased or induced by the compound(s), then the efficacy of the compound(s) may be decreased. For example, if the compound(s) increase or induce the expression of ABC B1 (MDR1), ABC C1 (MRP1), ABC C2 (MRP2), or ABC G2 (BCRP), then the efficacy of that compound may be lowered due to increased transport out of the cell. Conversely, if 15 the expression of one or more of the ABC transporter genes in the test sample is decreased or suppressed by the compound(s), then the efficacy and/or the toxicity of the compound(s) may be increased. For example, if the compound(s) decrease or suppress the expression of ABC B1 (MDR1), ABC C1 (MRP1), ABC C2 (MRP2), or ABC G2 (BCRP), then the efficacy and/or toxicity of that compound may be 20 increased due decreased transport out of the cell. This information is particularly important when designing drug treatments, including dosing amounts, for a particular disease.

In an embodiment of the invention, the compound is administered to a subject and ABC transporter gene expression is profiled in a test sample from the subject 25 before and/or after administration of the compounds. Changes in ABC transporter gene expression are indicative of the toxicity and/or efficacy of the compound in the subject. In a further embodiment, the subject is human.

In a further embodiment, the nucleic acids and methods of the present 30 invention are used to determine drug/drug interactions and their concomitant effect of ABC transporter gene expression. When two or more drugs are administered together, for example in combination therapy, ABC transporter gene expression may be altered. This is particularly relevant if two or more drugs are transported by the same transporter. What might be a non-toxic dose of a drug when administered on

its own, may turn into a toxic dose when that drug is administered along with another drug, for example if both drugs are substrates for the same transporter. Therefore it is important to determine a drug's effect on ABC transporter gene expression alone, as well as in the presence of one or more other drugs with which it may be co-administered. Accordingly, in a further embodiment of the present invention there is provided a method for determining a change in ABC transporter gene expression profile for a compound in the presence of one or more different compounds comprising:

- 10 (a) preparing an ABC transporter gene expression profile, using a method of the invention, of a test sample that has been exposed to the compound;
- 15 (b) preparing an ABC transporter gene expression profile, using a method of the invention, of a test sample that has been exposed to the compound and the one or more different compounds; and
- (c) quantitatively or qualitatively comparing the gene expression profile in (a) and (b),

wherein differential expression in (a) and (b) indicates that ABC transporter gene expression profile of the compound changes in the presence of the one or more different compounds.

In an embodiment of the invention, differential expression indicates the presence of drug-drug interactions. If drug-drug interactions are found, then caution would need to be taken when determining effective drug therapies, including dosing, when the drugs are to be present in the body or cell at the same time.

The methods of the present invention may also be used to monitor the changes in ABC transporter gene expression profile as a function of disease state. For example, an ABC transporter gene expression profile of a test sample from the subject may be obtained at one point in time and again at a later date. Changes in ABC transporter gene expression profile are indicative of changes in disease state, treatment response or treatment toxicity.

Another embodiment of the invention is the use of the ABC transporter gene expression information for population profiling. For example, the ABC transporter gene expression information can be used to pre-selected individuals for clinical trials into non-responder and responder groups to a particular drug or chemical before initiation of the clinical trial.

(VI) Databases

The present invention also includes relational databases containing ABC transporter gene expression profiles in various tissue samples and/or cell lines. The database may also contain sequence information as well as descriptive information about the gene associated with the sequence information, the clinical status of the test sample and/or its source. Methods of configuring and constructing such databases are known to those skilled in the art (see for example, Akerblom *et al.* 5,953,727).

The databases of the invention may be used in methods to identify the expression level in a test sample of the ABC transporter genes by comparing the expression level at least one of the ABC transporter genes in the test sample with the level of expression of the gene in the database. Such methods may be used to assess the physiological state or a given test sample by comparing the level of expression of an ABC transporter gene or genes in the sample with that found in samples from normal, untreated samples or samples treated with other agents.

(VII) Kits

The present invention further includes kits combining, in different combinations, nucleic acid arrays or microarrays, reagents for use with the arrays, signal detection and array-processing instruments, gene expression databases and analysis and database management software described above. The kits may be used, for example, to predict or model the toxic or therapeutic response of a test compound, to monitor the progression of disease states, to identify genes that show promise as new drug targets and to screen known and newly designed drugs as discussed above.

The databases packaged with the kits are a compilation of expression patterns from human or laboratory animal ABC transporter genes. Data is collected from a repository of both normal and diseased animal tissues and provides reproducible, quantitative results, i.e., the degree to which a gene is up-regulated or down-regulated under a given condition.

The kits may be used in the pharmaceutical industry, where the need for early drug testing is strong due to the high costs associated with drug development, but where bioinformatics, in particular gene expression informatics, is still lacking. These kits will reduce the costs, time and risks associated with traditional new drug

screening using cell cultures and laboratory animals. The results of large-scale drug screening of pre-grouped patient populations, pharmacogenomics testing, can also be applied to select drugs with greater efficacy and fewer side-effects. The kits may also be used by smaller biotechnology companies and research institutes who do not 5 have the facilities for performing such large-scale testing themselves.

Databases and software designed for use with use with microarrays is discussed in Balaban et al., U.S. Pat. No. Nos. 6,229,911, a computer-implemented method for managing information, stored as indexed tables, collected from small or large numbers of microarrays, and U.S. Pat. No. 6,185,561, a computer-based 10 method with data mining capability for collecting gene expression level data, adding additional attributes and reformatting the data to produce answers to various queries. Chee et al., U.S. Pat. No. 5,974,164, disclose a software-based method for identifying mutations in a nucleic acid sequence based on differences in probe fluorescence intensities between wild type and mutant sequences that hybridize to 15 reference sequences.

(VIII) Methods of Conducting Drug Discovery Businesses

Yet another aspect of the present invention provides a method of conducting a target discovery business comprising:

- (a) providing one or more assay systems for identifying agents by their 20 ability to modulate ABC transporter gene expression, said assay systems using a method of the invention;
- (b) (optionally) conducting therapeutic profiling of agents identified in step (a) for efficacy and toxicity in animals; and
- (c) licensing, to a third party, the rights for further drug development and/or 25 sales or agents identified in step (a), or analogs thereof.

By assay systems, it is meant, the equipment, reagents and methods involved in conducting a screen of compounds for the ability to modulate ABC transporter gene expression using the method of the invention.

The following non-limiting examples are illustrative of the present invention:

30 EXAMPLES

Example 1: Sets of primers and resulting PCR products for each ABC transporter gene

The sets of primers were designed such that the amplification product is a PCR amplicon that is a unique portion of an ABC transporter gene (See table 1). Figures 1 to 47 show nucleic acid sequences for each PCR amplicon. The primers are shown in bold.

5 The NCBI (www.ncbi.nlm.nih.gov) and BCM search launcher (www.searchlauncherbcm.tmc.edu) websites were used to verify PCR primer identity with the ABC transporter gene region of interest. BLAST sequence searches and alignment analyses were completed for each PCR primer pair and PCR amplicon to ensure minimum cross-hybridization with other known genes and other
10 known ABC transporter genes.

Total RNA preparation

Cell lines were grown as adherent monolayers following the ATCC guidelines in Falcon T175 flasks until semi-confluent. Culture medium was removed. The adherent cells were washed twice with PBS (phosphate buffered saline) pH7.4.
15 1.6ml TriZol reagent (Cat. No. 15596-018, Invitrogen Life Technologies) was added to each flask to lyse the cells and liberate the nucleic acids. The total RNA component of the nucleic acid lysate was isolated according to the manufacturer's instructions. Total RNA was quantitated by spectrophotometric analysis and OD_{260nm}:OD_{280nm} ratios.

cDNA synthesis

cDNA was prepared from 20 μ g of total RNA in a total volume of 40 μ l. 20 μ g of total RNA was added to a 200 μ l RNase-free microtube and placed on ice. 4 μ l of a 300ng/ μ l solution of random d(N)₉ primers (Cat. No. S1254S, New England BioLabs) was added to the tube containing the total RNA and the final volume made up to
20 22 μ l with RNase-free dH₂O. The microtube was capped and then heated at 65°C for 10min in a thermal cycler (PTC200 DNA Engine, MJ Research). The microtube was then removed from the thermal cycler and placed on ice for 3min. The microtube was spun in a microfuge (C-1200, VWR Scientific Products) to collect the solution in the bottom of the microtube and placed on ice.

0 First-strand cDNA synthesis was accomplished with the SuperScript II RNase H-Reverse Transcriptase reagent set (Cat. No. 18064-014, Invitrogen Life Technologies). 8ul 5x First-Strand Buffer [250mM Tris-HCl pH 8.3, 375mM KCl,

15mM MgCl₂], 4μl 100mM DTT, 2μl 10mM dNTP Mix [10mM each dATP, dCTP, dGTP, dTTP] were added to the microtube on ice. The microtube was capped and then heated at 25°C for 10min in a thermal cycler. The microtube was then heated at 42°C for 2min in a thermal cycler. The microtube was uncapped and left in the

5 thermal cycler. 2μl SuperScript II (200U/μl) was added to the solution in the microtube and mixed with the micropipette tip. The microtube was recapped and incubated at 42°C for 60min in a thermal cycler. Subsequent to this incubation the microtube was heated at 70°C for 15min in a thermal cycler. The microtube was then removed from the thermal cycler and spun in a microfuge to collect the solution in

10 the bottom of the microtube and then returned to the thermal cycler. 1μl of RNase H (2U/μl) was added to the cDNA synthesis reaction and incubated at 37°C for 20min in a thermal cycler. The first-strand cDNA synthesis reaction was then stored at -20°C until required for RT-PCR.

RT-PCR

15 RT-PCR was performed in a final volume of 25μl. 2μl of the first-strand cDNA synthesis reaction was added to a 200μl microtube and placed on ice. 2μl of a specific ABC Drug Transporter (ABC-DT) primer pair mix [10μM each forward PCR primer and reverse PCR primer], 2.5μl 10x PCR Buffer [200mM Tris-HCl pH 8.4, 500mM KCl], 0.75μl 50mM MgCl₂, 0.5μl 10mM dNTP Mix [10mM each dATP, dCTP, dGTP, dTTP], 16.25μl dH₂O and 1μl Taq polymerase (5U/μl) were added to the side of the microtube. The reagents were mixed and collected in the bottom of the microtube by spinning the capped microtube in a microfuge. The capped microtube was then placed in a thermal cycler block with a heated lid (PTC200 DNA Engine, MJ Research), both pre-heated to 95°C, and incubated at this temperature for 5min.

20

25 After this initial denaturation step 40 cycles of PCR amplification were performed as follows: Denature 95°C for 30s, Anneal 60°C for 30s, Extend 72°C for 60s. Following the final 72°C Extend step the PCR was incubated for an additional 10min at 72°C. The PCR was then maintained at a temperature of 15°C. PCR products were stored at -20°C until needed.

30 PCR amplicon purification

ABC-DT RT-PCR amplification products (PCR amplicons) were analysed by electrophoresis at 150V for 20min in 1x TAE running buffer in an agarose gel [0.8%

agarose, 1x TAE, 0.5 μ g/ml ethidium bromide] with 4 μ l of a 250bp DNA Ladder (Cat. No. 10596-013, Invitrogen Life Technologies) to permit size estimates of the PCR amplicons.

5 The ABC-DT RT-PCR amplification products (PCR amplicons) were visualised "in gel" with a UV transilluminator (UVP M-15, DiaMed Lab Supplies) and photographed with a photo-documentation camera and hood (FB-PDC-34, FB-PDH-1216, Fisher Biotech), a #15 Deep Yellow 40.5mm screw-in optical glass filter (FB-PDF-15, Fisher Biotech) and Polaroid Polapan 667 film.

10 The ABC-DT RT-PCR amplification products (PCR amplicons) were isolated and purified from the ABC-DT RT-PCR using the QIAquick PCR purification kit (Cat. No. 28104, QIAGEN Inc.) according to the manufacturer's instructions. After purification, ABC-DT RT-PCR amplification products (PCR amplicons) were analysed by electrophoresis at 150V for 20min in 1x TAE running buffer in an agarose gel [0.8% agarose, 1x TAE, 0.5 μ g/ml ethidium bromide] with 4 μ l of a Low 15 DNA Mass Ladder (Cat. No. 10068-013, Invitrogen Life Technologies) to permit PCR amplicon sizing and quantitation.

20 Figure 48 shows the ABC transporter gene RT-PCR amplification products from the CaCo2 cell line. Figure 49 shows the ABC transporter gene RT-PCR amplification products from the HEK293 cell line. Figure 50 shows the ABC transporter gene RT-PCR amplification products from the HepG2 cell line.

Example 2: Sequencing

The sequences of the PCR amplicons, which are each unique portions of each of the known human ABC transporter genes, can be verified.

ABC-DT PCR amplicon cloning and sequencing

25 A number of the purified ABC-DT RT-PCR amplification products (PCR amplicons) were cloned into pCR4-TOPO vectors using the TOPO TA Cloning Kit for Sequencing (Cat. No. K4575-40, Invitrogen Life Technologies) according to the manufacturer's instructions to verify the sequence of the purified ABC-DT PCR amplicon.

0 DNA sequence analysis was performed with Cy5.5-labelled M13 (-20) universal and M13 reverse primers, the Cy5/Cy5.5 Dye Primer Cycle Sequencing Kit (Cat. No. VG 30001, Visible Genetics Inc./Bayer Inc.) and the OpenGene automated

DNA sequencing system (MGB-16, Visible Genetics Inc./Bayer Inc.) according to the manufacturer's instructions.

Example 3: DNA Microarray

ABC-DT microarray (DT1 microarray)

5 1-2 μ g of each of the purified ABC-DT RT-PCR amplification products (PCR amplicons) and 5 purified positive control RT-PCR amplification products (PCR amplicons) were aliquoted into individual wells of a CoStar SeroCluster 96 well U-bottom polypropylene microwell plate (source plate). The source plate was placed in a Speed-Vac concentrator (SPD101B, Savant Instruments Inc.) and dried under
10 vacuum for 1 hour at 45°C. The dry RT-PCR amplification products (PCR amplicons) in the source plate were resuspended in 20 μ l 1x NoAb Print Buffer (150mM sodium phosphate pH 8.5, Cat. No. UAS0001PB, NoAb BioDiscoveries Inc.), sealed with mylar sealing tape (Cat. No. T-2162, Sigma Chemical Company) and dissolved by shaking at 300rpm for 1 hour at room temperature on a microplate shaker (EAS2/4,
15 SLT Lab Instruments).

The source plate was then placed in a humidified (21-25°C, 45-60% RH) microarrayer cabinet (SDDC-2, ESI / Virtek Vision Corp. / BioRad Laboratories Inc.). Each purified RT-PCR amplification product (PCR amplicon) was printed in quadruplicate on activated covalent-binding epoxy slides (Cat. No. UAS0005E, NoAb
20 BioDiscoveries Inc.) using Stealth micro-spotting pins (Cat. No. SMP5, TeleChem International Inc.). The 384 element microarrays were air-dried in the microarrayer cabinet for at least 4 hours. Printed microarrays were stored in 20 slide racks under vacuum until needed.

**Example 4: Method for detecting ABC transporter gene expression using a
25 DNA microarray**

The ABC transporter gene expression profile for 22 different cell lines was prepared using the DNA microarray.

Total RNA preparation

All 22 cell lines (BT20, CaCo2, CaOv, Colo320, HBT161, HEK293, HepG2,
30 HT75, HT177, LnCaP, MCF7, MDA453, MDA468, MFE29C, SKMES1, SKNAS, SKNBE, SKND2, SKNMC, T47D, ZR75, MDCK) were grown as adherent monolayers following the ATCC guidelines in tissue culture flasks until semi-confluent. Culture medium was removed. The adherent cells were washed twice with

PBS (phosphate buffered saline) pH7.4. 1.6ml Trizol reagent (Cat. No. 15596-018, Invitrogen Life Technologies) was added to each flask to lyse the cells and liberate the nucleic acids. The total RNA component of the nucleic acid lysate was isolated according to the manufacturer's instructions. Total RNA was quantitated by 5 spectrophotometric analysis and OD_{260nm}:OD_{280nm} ratios.

Fluorescent cDNA target preparation

Fluorescently labelled cDNA targets were prepared from each of the 22 cell lines using 20 μ g of total RNA in a total volume of 40 μ l.

20 μ g of total RNA was added to a 200 μ l RNase-free microtube and placed on 10 ice. 4 μ l of a 300ng/ μ l solution of random d(N)₉ primers (Cat. No. S1254S, New England BioLabs) was added to the tube containing the total RNA and the final volume made up to 22 μ l with RNase-free dH₂O. The microtube was capped and then heated at 65°C for 10min in a thermal cycler (PTC200 DNA Engine, MJ Research). The microtube was then removed from the thermal cycler and placed on ice for 3min. 15 The microtube was spun in a microfuge (C-1200, VWR Scientific Products) to collect the solution in the bottom of the microtube and placed on ice.

First-strand cDNA synthesis was accomplished with the SuperScript II RNase H-Reverse Transcriptase reagent set (Cat. No. 18064-014, Invitrogen Life Technologies). 8 μ l 5x First-Strand Buffer [250mM Tris-HCl pH 8.3, 375mM KCl, 20 15mM MgCl₂], 4 μ l 100mM DTT, 2 μ l T- dNTP Mix [2.3mM dTTP, 5mM each dATP, dCTP, dGTP], 2 μ l ChromaTide Alexa 546-14-dUTP (1mM in TE buffer, Cat. No. C-11401, Molecular Probes Inc.) were added to the microtube on ice. The microtube was capped and then heated at 25°C for 10min in a thermal cycler. The microtube was then heated at 42°C for 2min in a thermal cycler. The microtube was uncapped 25 and left in the thermal cycler. 2 μ l SuperScript II (200U/ μ l) was added to the solution in the microtube and mixed with the micropipette tip. The microtube was recapped and incubated at 42°C for 60min in a thermal cycler. Subsequent to this incubation the microtube was heated at 70°C for 15min in a thermal cycler. The microtube was then removed from the thermal cycler and spun in a microfuge to collect the solution 30 in the bottom of the microtube and then returned to the thermal cycler. 1 μ l of RNase H (2U/ μ l) was added to the cDNA synthesis reaction and incubated at 37°C for 20min in a thermal cycler. The fluorescently labelled cDNA targets were stored at

-20°C overnight before QIAquick column purification.

The fluorescantly labelled cDNA targets were thawed and the total volume adjusted to 100µl with dH₂O. Labelled cDNA targets were isolated and purified using the QIAquick PCR purification kit (Cat. No. 28104, QIAGEN Inc.) according to the 5 manufacturer's instructions except that the final elution volume was adjusted to 150µl. The purified cDNA target preparation was stored at -20°C until required for microarray hybridisation.

DT1 microarray hybridisation

The printed DT1 microarray(s) was removed from storage under vacuum and 10 placed in a 20 slide rack. The DT1 microarray was then denatured by dipping the microarray slide into "boiled" dH₂O for 30s. The denatured DT1 microarray was then placed in a polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) and blocked in 1x NoAb Pre-Hybridisation Blocking Buffer (Cat. No. UAS0001BB, NoAb BioDiscoveries Inc.) for 2 hours at room temperature. Pre-15 hybridised, blocked DT1 microarrays were removed from this solution and placed in a new polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) containing a solution of denatured, labelled cDNA targets from a specific cell line.

The labelled cDNA target preparation was thawed and the 150µl added to 850µl hybridisation buffer (500mM sodium Phosphate pH 6.0, 1% SDS, 1% BSA, 20 1mM EDTA) in a 1.5ml microtube and heated at 95°C for 10min. Following denaturation the microtube was spun briefly in a microcentrifuge to collect all the liquid. The denatured, labelled cDNA targets were then added to a polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) that contained a pre-hybridised, blocked DT1 microarray placed "array-side" down in the bottom-most slot 25 of the 5 slide mailer. In this orientation the entire surface of the microarray slide is bathed in the hybridisation buffer. 5 slide mailers containing the DT1 microarrays were incubated on their sides, "array-side" down, in a 37°C incubator for 15-18h.

Hybridised DT1 microarrays were removed from the 5 slide mailers with 30 forceps and placed directly into a 20 slide rack in a slide wash box containing a 0.1x SSC, 0.1% SDS solution. DT1 microarrays were incubated in this solution at 37°C for 15min. The slide rack containing the DT1 microarrays was then transferred to a slide wash box containing 0.1x SSC and incubated in this solution at 37°C for 15min.

Following this step the DT1 microarrays were rinsed in dH₂O and air-dried by centrifugation at 1200rpm.

DT1 microarray image acquisition and data analysis

Processed DT1 microarrays were scanned using ScanArray software in a 5 ScanArray Lite MicroArray Analysis System (GSI Lumonics Inc.) at a scan resolution of 10µm, a laser setting of 90 and a PMT gain of 80. Images were analysed using QuantArray software (GSI Lumonics Inc.). The data generated from QuantArray was exported to GeneLinker Gold (Molecular Mining Inc. / Predictive Patterns Software) for bioinformatic analysis and data mining. Gene expression profiles and hierarchical 10 clustering maps ("heat maps") were also generated using GeneLinker Gold.

Figure 51 shows the fluorescence intensity cluster plot for and Table 2 sets out the relative levels of ABC transporter gene expression in various cell lines normalized to GAPDH. Figure 52 shows the fluorescence intensity cluster plot for and Table 3 sets out the relative levels of ABC transporter gene expression in 15 various cell lines normalized to actin. Figure 53 shows the fluorescence intensity cluster plot for and Table 4 sets out the relative levels of ABC transporter gene expression in various cell lines normalized to SH1.

Figure 54 shows the relative levels of gene expression for ABC B1 to B11 in HEK cells normalized to constitutively expressed control genes (tubulin, actin, 10 GAPDH, and SH1). Figure 55 shows the relative levels of gene expression for ABC B1 to B11 in various cell lines (HEK, CaCo2, CaOv and HepG2) normalized to the constitutively expressed actin control gene.

As shown in Figure 55, the ABC transporter gene expression profile is different for different cell lines. Certain ABC transporter genes are over-expressed in 5 some cell lines, while some are suppressed in other cell lines.

Example 5: Drug screening assay

Cell lines were treated with two chemotherapeutic agents, doxorubicin and vinblastine, at 2 hour intervals.

Total RNA preparation from drug-treated HepG2 cell line

0 The HepG2 cell line was grown as an adherent monolayer in 24 Falcon T175 flasks following the ATCC guidelines until semi-confluent. Tissue culture flasks were then divided into pairs for each of six timepoints (0h, 2h, 4h, 8h, 18h, 24h).

For vinblastine sulfate treatment, 5 μ l of a 1000x (5mM in DMSO) stock solution of vinblastine sulfate was added to 10 Falcon T175 flasks containing the HepG2 monolayer in 10mls of culture medium (25nM final concentration), mixed gently by rocking, returned to the CO₂ incubator and harvested for total RNA at the 5 indicated times. The 0h timepoint flasks were processed immediately after the addition of 5 μ l DMSO.

For doxorubicin HCl treatment, 5 μ l of a 1000x (5mM in DMSO) stock solution of doxorubicin HCl was added to 10 Falcon T175 flasks containing the HepG2 monolayer in 10mls of culture medium (25nM final concentration), mixed gently by 10 rocking, returned to the CO₂ incubator and harvested for total RNA at the indicated times. The 0h timepoint flasks were processed immediately after the addition of 5 μ l DMSO.

Prior to cell lysis the tissue culture medium was removed. The adherent cells were washed twice with PBS (phosphate buffered saline) pH7.4. 1.6ml TriZol 15 reagent (Cat. No. 15596-018, Invitrogen Life Technologies) was added to each flask to lyse the cells and liberate the nucleic acids. The total RNA component of the nucleic acid lysate was isolated according to the manufacturer's instructions. Total RNA was quantitated by spectrophotometric analysis and OD_{260nm}:OD_{280nm} ratios.

Fluorescent cDNA target preparation

20 Fluorescently labelled cDNA targets were prepared from each of the 12 timepoint samples for the drug-treated HepG2 cell line (6x vinblastine sulfate, 6x doxorubicin HCl) using 20 μ g of total RNA in a total volume of 40 μ l.

25 20 μ g of total RNA was added to a 200ul RNase-free microtube and placed on ice. 4 μ l of a 300ng/ μ l solution of random d(N)₉ primers (Cat. No. S1254S, New England BioLabs) was added to the tube containing the total RNA and the final volume made up to 22 μ l with RNase-free dH₂O. The microtube was capped and then heated at 65°C for 10min in a thermal cycler (PTC200 DNA Engine, MJ Research). The microtube was then removed from the thermal cycler and placed on ice for 3min. The microtube was spun in a microfuge (C-1200, VWR Scientific Products) to collect 30 the solution in the bottom of the microtube and placed on ice.

First-strand cDNA synthesis was accomplished with the SuperScript II RNase H- Reverse Transcriptase reagent set (Cat. No. 18064-014, Invitrogen Life

Technologies). 8 μ l 5x First-Strand Buffer [250mM Tris-HCl pH 8.3, 375mM KCl, 15mM MgCl₂], 4 μ l 100mM DTT, 2 μ l T- dNTP Mix [2.3mM dTTP, 5mM each dATP, dCTP, dGTP], 2 μ l ChromaTide Alexa 546-14-dUTP (1mM in TE buffer, Cat. No. C-11401, Molecular Probes Inc.) were added to the microtube on ice. The microtube
5 was capped and then heated at 25°C for 10min in a thermal cycler. The microtube was then heated at 42°C for 2min in a thermal cycler. The microtube was uncapped and left in the thermal cycler. 2 μ l SuperScript II (200U/ μ l) was added to the solution in the microtube and mixed with the micropipette tip. The microtube was recapped and incubated at 42°C for 60min in a thermal cycler. Subsequent to this incubation
10 the microtube was heated at 70°C for 15min in a thermal cycler. The microtube was then removed from the thermal cycler and spun in a microfuge to collect the solution in the bottom of the microtube and then returned to the thermal cycler. 1 μ l of RNase H (2U/ μ l) was added to the cDNA synthesis reaction and incubated at 37°C for 20min in a thermal cycler. The fluorescently labelled cDNA targets were stored at
15 -20°C overnight before QIAquick column purification.

The fluorescently labelled cDNA targets were thawed and the total volume adjusted to 100 μ l with dH₂O. Labelled cDNA targets were isolated and purified using the QIAquick PCR purification kit (Cat. No. 28104, QIAGEN Inc.) according to the manufacturer's instructions except that the final elution volume was adjusted to
20 150 μ l. The purified cDNA target preparation was stored at -20°C until required for microarray hybridisation.

DT1 microarray hybridisation

The printed DT1 microarray(s) was removed from storage under vacuum and placed in a 20 slide rack. The DT1 microarray was then denatured by dipping the
25 microarray slide into "boiled" dH₂O for 30s. The denatured DT1 microarray was then placed in a polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) and blocked in 1x NoAb Pre-Hybridisation Blocking Buffer (Cat. No. UAS0001BB, NoAb BioDiscoveries Inc.) for 2 hours at room temperature. Pre-hybridised, blocked DT1 microarrays were removed from this solution and placed in
30 a new polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) containing a solution of denatured, labelled cDNA targets from a specific cell line.

The labelled cDNA target preparation was thawed and the 150µl added to 850ul hybridisation buffer (500mM sodium Phosphate pH 6.0, 1% SDS, 1% BSA, 1mM EDTA) in a 1.5ml microtube and heated at 95°C for 10min. Following denaturation the microtube was spun briefly in a microcentrifuge to collect all the 5 liquid. The denatured, labelled cDNA targets were then added to a polypropylene 5 slide mailer (Cat. No. 240-3074-030, Evergreen Scientific) that contained a pre-hybridised, blocked DT1 microarray placed "array-side" down in the bottom-most slot of the 5 slide mailer. In this orientation the entire surface of the microarray slide is bathed in the hybridisation buffer. 5 slide mailers containing the DT1 microarrays 10 were incubated on their sides, "array-side" down, in a 37°C incubator for 15-18h.

Hybridised DT1 microarrays were removed from the 5 slide mailers with forceps and placed directly into a 20 slide rack in a slide wash box containing a 0.1x SSC, 0.1% SDS solution. DT1 microarrays were incubated in this solution at 37°C for 15min. The slide rack containing the DT1 microarrays was then transferred to a 15 slide wash box containing 0.1x SSC and incubated in this solution at 37°C for 15min. Following this step the DT1 microarrays were rinsed in dH₂O and air-dried by centrifugation at 1200rpm.

DT1 microarray image acquisition and data analysis

Processed DT1 microarrays were scanned using ScanArray software in a 20 ScanArray Lite MicroArray Analysis System (GSI Lumonics Inc.) at a scan resolution of 10µm, a laser setting of 90 and a PMT gain of 80. Images were analyzed using QuantArray software (GSI Lumonics Inc.). The data generated from QuantArray was exported to GeneLinker Gold (Molecular Mining Inc. / Predictive Patterns Software) for bioinformatic analysis and data mining. Gene expression profiles and hierarchical 25 clustering maps for drug treatment-related changes in ABC-DT gene expression were also generated using GeneLinker Gold.

Figure 56 shows the fluorescence intensity cluster plot for and Table 5 shows the relative levels of ABC transporter gene expression in cell lines treated with doxorubicin at various time intervals. Figure 57 shows the fluorescence intensity 30 cluster plot for and Table 6 shows the relative levels of ABC transporter gene expression in cell lines treated with vinblastine at various time intervals.

Figure 58 shows a matrix plot of the relative levels of ABC transporter gene expression in a cell line [HepG2] treated with either doxorubicin [dox] or vinblastine [vin] at various time intervals.

Figure 59 shows a matrix plot of the relative levels of ABC transporter gene

5 expression in several cell lines [A549, CaCo2, HepG2] treated with either acetaminophen [AP] or acetylsalicylic acid [SA].

Figure 60 shows a matrix plot of the relative levels of ABC transporter gene expression in a cell line [A549] treated with either all-trans retinoic acid [AAT], cis-13 retinoic acid [A13], cis-9 retinoic acid [A9] or phorbol-12-myristate-13-acetate [APM].

10 Figure 61 shows a matrix plot of the relative levels of ABC transporter gene expression in cell lines HTB81 [A], CRL1740 [C] and CRL2505 [D] treated with either no drug [none], methanol [Me], phenobarbital [PhB], acetylsalicylic acid [ASA] or acetaminophen [AAP].

15 While the present invention has been described with reference to what are presently considered to be examples, it is to be understood that the invention is not limited to the disclosed examples. To the contrary, the invention is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the appended claims.

20 All publications, patents and patent applications are herein incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

Table 1

Unique Portion of ABC Transporter Gene		Upper Primer		Lower Primer
ABCA1	SEQ ID NO: 48	5' CCC TGT GGA ATG TAC CTA TGT GAG 3'	SEQ ID NO: 49	5' GCG TAA AGT GCT TGG AAT GAG GGC 3'
ABCA2	SEQ ID NO: 50	5' CCT TCA ACA CGG ACA CGC TCT GCT 3'	SEQ ID NO: 51	5' AGC TTC TCC ATT CCT GCC ACC TGC 3'
ABCA3	SEQ ID NO: 52	5' AAG GAA AAG TAC GGC GTG GAC GAC 3'	SEQ ID NO: 53	5' CTA AGA CCC CAG CAC CTA ATC ACA 3'
ABCA4	SEQ ID NO: 54	5' GAG CAT CAT CAG AAA AGG GAG GGC 3'	SEQ ID NO: 55	5' GGG TTT CTA GTT CTG GGG TCT GGA 3'
ABCA5	SEQ ID NO: 56	5' AAT GCA AGC CGT CAG GAA AGT TTT 3'	SEQ ID NO: 57	5' CTT ACA CTT CAG CTT TTA CGG ATG 3'
ABCA6	SEQ ID NO: 58	5' AGT TGT GTT TTG TGC TGA GCC TCC 3'	SEQ ID NO: 59	5' GTG CCT GAC TCT TTG GGT GAC TTT 3'
ABCA7	SEQ ID NO: 60	5' ATA GCA TGG AGG AGT GTG AAG CGC 3'	SEQ ID NO: 61	5' TTT CAC CAC CAC GGC TTC TCT CCA 3'
ABCA8	SEQ ID NO: 62	5' GCT GGG TGA TTT TGA GGA GGA TTT 3'	SEQ ID NO: 63	5' GAA AAT GGC ACA CAG TTG GCT TAC 3'
ABCA9	SEQ ID NO: 64	5' TGT GCC AGC AAC CAA ATC CCA TGT 3'	SEQ ID NO: 65	5' TTT CTC CTA ATG CTA TCC CTC CCC 3'
ABCA10	SEQ ID NO: 66	5' AGG AGC TGG GAA ATG TTG ATG ATA 3'	SEQ ID NO: 67	5' GCC ATT TCA TCA GTT TAT CAG ACC 3'
ABCA12	SEQ ID NO: 68	5' CCT GCT GGA GAG TGT TTT GGG CTT 3'	SEQ ID NO: 69	5' ATG TTT GCG ACT CCT CCT GCT GTG 3'
ABCB1	SEQ ID NO: 70	5' CAT CCT GTT TGA CTG CAG CAT TGC 3'	SEQ ID NO: 71	5' GCA AGG CAG TCA GTT ACA GTC CAA 3'
ABCB2	SEQ ID NO: 72	5' ATA TTG CCT ATG GCC TGA CCC AGA 3'	SEQ ID NO: 73	5' TTC TCA GTT TCA GAG TGC TGG CCA 3'
ABCB3	SEQ ID NO: 74	5' GGG AGT AGG AGC TAT GCT AAG TGT 3'	SEQ ID NO: 75	5' TGC TCA TGG TCT AGT GGA AGG TCA 3'
ABCB4	SEQ ID NO: 76	5' TTG ACA GCT ACA GTG AAG AGG GGC 3'	SEQ ID NO: 77	5' CAT AAG TTC TGT GTC CCA GCC TGG 3'
ABCB6	SEQ ID NO: 78	5' TTC GCT TCT ACG ACA TCA GCT CTG 3'	SEQ ID NO: 79	5' GAC CAG GAT GAA ATA AGC CAG GGA 3'
ABCB7	SEQ ID NO: 80	5' CCC TGC	SEQ ID NO: 81	5' CTT AGC ACG

		AGG AAA GAA AGT GGC CAT 3'		AAC AGT TTC CAC AGC 3'
ABCB8	SEQ ID NO: 82	5' AGG TTG TCG GTT TCA TCA GCC AGG 3'	SEQ ID NO: 83	5' TTT ATT GTG AGC AGG AGC AGC CGC 3'
ABCB9	SEQ ID NO: 84	5' TGG ATC ACC GCT TCC TGC ATC TTG 3'	SEQ ID NO: 85	5' TGC CAC CAT CCC ATC CAC CAA AGA 3'
ABCB10	SEQ ID NO: 86	5' GCA AGG CAT GAA CTG CTA GGT ATT 3'	SEQ ID NO: 87	5' GGT TTC TTC TTC CAG TCT AAT CAG 3'
ABCB11	SEQ ID NO: 88	5' TTG TCA TTG CCC ATC GCT TGT CCA 3'	SEQ ID NO: 89	5' AGA GCA TCC ACC CTT TCC CTA TCC 3'
ABCC1	SEQ ID NO: 90	5' GCT CCC ATC ACC TCT AAC ATC CTT 3'	SEQ ID NO: 91	5' TGA GCA GGT ACC ATG AGA GGG AAA 3'
ABCC2	SEQ ID NO: 92	5' GTA GCA TGG AGA AGA TTG GTG TGG 3'	SEQ ID NO: 93	5' GGG TAG TAG GTT CAT GGG TGT TCA 3'
ABCC3	SEQ ID NO: 94	5' CAA GAG CCG CAT CCT GGT TTT AGA 3'	SEQ ID NO: 95	5' TTT AAT GGA TTC AGG CAG CAC CCC 3'
ABCC4	SEQ ID NO: 96	5' TGG GAA GAA CCG GAG CTG GAA AAA 3'	SEQ ID NO: 97	5' AAT GCC TTC GGA ACG GAC TTG ACA 3'
ABCC5	SEQ ID NO: 98	5' AAG GAA GAC GTG TGG CAA TAG TGG 3'	SEQ ID NO: 99	5' AAA CCA CAC AGC AAC CAG CAA CCT 3'
ABCC6	SEQ ID NO: 100	5' TCG TGT CAG TGG AGC GGA TGC AGG 3'	SEQ ID NO: 101	5' CTG CCA CCT GCC CCT TGT CCA TGA 3'
ABCC7	SEQ ID NO: 102	5' TCT TTC ACA GGG GAC AGG ATG GTT 3'	SEQ ID NO: 103	5' CAG TTT GGA GTT GAG AAG GCA GTG 3'
ABCC8	SEQ ID NO: 104	5' AAA CCG AGG CAG AGA GCT ACG AGG 3'	SEQ ID NO: 105	5' TGG GCT CTG GCA GGT CAC TTG TCT 3'
ABCC9	SEQ ID NO: 106	5' TGG GTG CAG TGA AGA AGG TGA ACA 3'	SEQ ID NO: 107	5' GTG GGC GAA CAA ATT TGG GAC AGT 3'
ABCC10b	SEQ ID NO: 108	5' TCT TCC CTG TTG TTG GTG CTC TTC 3'	SEQ ID NO: 109	5' TGA AAA TGC AAG TGG GCT CCT ATG 3'
ABCC11	SEQ ID NO: 110	5' GAT TCT CAT TGA CGG CGT GGA CAT 3'	SEQ ID NO: 111	5' TGG TTC TGG GGT TCT AAG GTC TTG 3'
ABCC12a	SEQ ID NO: 112	5' CTG GTT ATG GAA AAT GGG AAG GTG 3'	SEQ ID NO: 113	5' TTG CAA GGC GAC ATT TCA GGG TAA 3'
ABCC13	SEQ ID NO: 114	5' GCA CCT GTG GGC CAT ACT AAA AGA 3'	SEQ ID NO: 115	5' TAA CAA ACA CAA GGA CTG CCA CCC 3'
ABCD1	SEQ ID NO:	5' TTC CCT CCT	SEQ ID NO:	5' TCT TTG GCA

	116	CGT CAG TCT CTC AAA 3'	117	CTG AGC TGG GAA CAT 3'
ABCD2	SEQ ID NO: 118	5' GTG GCC AAC TAA ACC TGT ACA AAA 3'	SEQ ID NO: 119	5' ACA AAA GAG CAC TAA ACC AGA GAG 3'
ABCD3	SEQ ID NO: 120	5' TAC TCA TTC CTT GTG TGT GTC TTG 3'	SEQ ID NO: 121	5' CTT CGG TAG CCA GTG ATT GTT ATA 3'
ABCD4	SEQ ID NO: 122	5' CTC CAT ATG CTT GAA GTG CTG ATT 3'	SEQ ID NO: 123	5' AGA AGC CTG GCA AAC ATT ATG AAG 3'
ABCE1	SEQ ID NO: 124	5' ATT CCC CGC AAA AAA CCC CTA ACT 3'	SEQ ID NO: 125	5' TGG GAG GGT AAT AAA GGG AGA TCA 3'
ABCF1	SEQ ID NO: 126	5' TTG GAG GCC CTG GGT GAA GTC ATG 3'	SEQ ID NO: 127	5' TTT CCT GCC CCA AGT CCT CAA CCA 3'
ABCF2	SEQ ID NO: 128	5' TGC TAC CCA GAG ATC AAG GAG AAG 3'	SEQ ID NO: 129	5' ACT TGG AGC TGG TGT ACT TGG TGA 3'
ABCF3	SEQ ID NO: 130	5' CCT AAA CGT CAG TGC TTG TGG AAC 3'	SEQ ID NO: 131	5' TTT ACA TAG CAG CCA CTT GGG GTC 3'
ABCG1	SEQ ID NO: 132	5' CGT CTA GAA TCG AGG AGG CAA GCC 3'	SEQ ID NO: 133	5' CCA GCT GGG TGA CTC GGG TTA AAC 3'
ABCG2	SEQ ID NO: 134	5' CAG TAC TTC AGC ATT CCA CGA TAT 3'	SEQ ID NO: 135	5' GGG CTA CTA ACC TAC CTA TTC ATT 3'
ABCG4	SEQ ID NO: 136	5' ACA GGC ACA TAC ATG AGA ACA GGC 3'	SEQ ID NO: 137	5' CAG GGA TGT GTA CAG GAA AAA GGG 3'
ABCG5	SEQ ID NO: 138	5' GCC CAG GTG CAA CAT CTA GAT TCA 3'	SEQ ID NO: 139	5' CCC TCG TGT GGA CAT CTG CAT TTA 3'
ABCG8	SEQ ID NO: 140	5' TCA ATG ACC ATC GGC TTC CTC TAT 3'	SEQ ID NO: 141	5' ACG TAG TAC AGG ACC ATG AAG CCA 3'

Table 2

	h20	cac02	caov	col0320	hbt161	hek	hek2	hekG2	h75	h177	Incp	mc7	mda468	mfec9c	skms1	skms1	skbe	skbd2	skmc	tf7d	xt75	ndck				
w1	1.760147	1.618089	1.424148	1.161262	1.543551	1.987004	1.269118	1.040183	1.176464	1.602791	1.511799	1.655671	1.172517	1.3744591	1.257281	2.173767	0.626002	1.2967551	1.626099	1.447571	1.850718	1.613761	1.412566			
abcA1	3.047933	3.347099	0.8287261	2.370672	5.264526	3.609649	2.229297	1.564597	2.908955	4.401204	5.121514	3.141233	1.70476	3.2744642	3.10138	2.974131	1.551803	2.454231	3.391316	2.45395	2.551371	2.455901				
abcA2	4.624323	5.261656	2.509857	4.162548	7.839229	6.170351	4.6106237	2.813327	3.969959	6.672095	6.6802529	5.119029	3.48243	4.697005	4.960736	4.745097	1.381523	3.811958	5.770854	4.222122	6.598368	3.150876	3.995495			
sh1	3.911999	4.197328	5.598497	3.81232	6.310507	4.454382	4.764957	5.265198	3.87439	5.271368	6.399399	4.016223	2.307139	4.61252	3.6699887	3.949923	1.082382	3.228175	4.988276	3.187279	5.914972	2.348161	4.966444			
abcA3	7.608692	8.546286	4.317059	7.302629	10.71203	8.577419	6.242746	4.433605	6.165651	12.294561	11.60584	7.93316	5.148105	6.795211	7.890175	7.284888	2.99219	6.388617	9.331218	6.042775	9.776199	3.432724	5.13229			
abcA4	5.519356	7.41871	4.700107	6.432769	7.527029	8.965933	7.075456	4.358784	4.721274	8.165924	8.144223	6.643413	4.827968	5.660874	5.827968	5.660874	8.173683	6.244308	8.173683	5.636549	5.248557					
abcA5	2.957649	3.448256	0.89617	2.696655	4.770988	3.656885	4.230468	2.073816	3.708588	5.477282	3.097722	1.716878	4.339422	2.2885842	3.511156	1.228945	2.2581133	3.774496	2.166136	5.289771	2.315449	3.184366				
abcA6	5.16411	5.995906	2.218216	6.488178	7.475329	7.827521	6.676626	3.339979	4.716604	7.380911	8.603165	5.66999	4.405	5.3359	5.705216	6.032734	1.482493	4.932076	6.6565578	4.76329	7.771708	3.71513	4.6072			
abcA7	0.815024	1.451727	0.422628	0.756805	0.925006	1.414485	1.045833	1.075277	1.307867	1.28646	1.089256	0.780969	0.415919	1.64963	0.2951143	1.18934	1.18934	1.169245	0.678497	0.0242581	0.687287	1.060294	0.6227354	0.031147		
abcA8	3.766634	4.410631	1.384956	4.646162	5.742969	4.997531	3.428515	2.963395	3.964944	5.261313	6.470901	4.355213	2.630278	4.403083	3.3656871	4.097393	1.461138	3.412184	4.4223174	7.010437	6.046554	2.9000982	3.329838			
abcA9	6.74465357	7.8545984	2.973699	10.559248	11.38119	10.562825	10.784515	6.514559	6.197568	12.382926	13.074749	7.775526	6.282562	7.08791	8.729474	6.840259	1.466374	8.409215	11.61034	6.671209	10.10557	4.556159	7.380374			
abcA10	3.731852	4.922754	0.973826	3.084834	7.438836	4.674079	5.928152	8.11801	3.495781	8.054075	8.295051	3.571341	2.8155333	4.373392	4.0444166	4.283748	4.0745729	4.4228018	4.4554904	5.338708	6.187728	3.136154	5.292145			
abcA12	4.602116	6.046777	2.419136	6.102304	6.873125	5.533879	5.779729	3.000348	3.831084	7.761779	7.458226	5.19048	3.419758	4.452229	5.750435	4.411436	1.072138	4.130774	6.437921	5.93261	4.437921	5.922869				
abcB1	6.229988	6.922212	3.192804	6.876342	8.412035	7.777775	6.954351	2.88971	4.556779	9.103438	9.470861	6.403695	4.676444	5.728899	6.192011	6.088017	6.692989	5.384571	8.059313	5.604886	8.288023	5.028871	6.296814			
abcB2	4.692122	5.67407	2.941921	4.322886	7.075056	6.906656	5.510521	3.107792	4.274465	6.468082	6.484062	5.198367	3.292553	5.0566685	4.284439	4.6555056	2.352081	3.636367	8.826283	3.515015	6.949947	5.5383017	5.0333			
actin	2.813648	2.514351	2.139753	2.901062	2.465122	3.19905	3.166989	1.849321	2.14277	4.099057	2.0619	2.663159	2.310214	2.052918	2.5438381	2.572973	1.295335	2.524054	4.114827	2.527293	5.127294	2.1447456	2.8423669			
abcB3	5.815124	6.383326	4.205251	6.253251	8.191762	8.021734	8.277449	3.410767	5.018789	8.114998	7.876121	6.406229	4.4747808	6.190622	4.999937	6.749297	2.140021	5.4338017	9.335644	3.940163	8.353189	4.062514	7.246606			
abcB4	7.6835	8.38654	7.058921	10.66992	10.24258	12.0061	12.24046	6.266877	6.656844	12.67506	11.05904	8.330893	6.509777	7.778177	7.761991	7.427915	8.12579	8.712528	14.492689	7.130204	10.49239	5.045277	7.209005			
abcB5	4.084981	4.984172	2.846189	4.31622	5.588544	5.753428	5.929129	2.89158	4.190264	6.217372	5.877825	4.392925	2.776184	4.6565543	4.099493	4.31774	1.576024	3.79744	1.576024	3.79744	1.576024	3.1386715	3.3186715			
abcB7	0.074123	0.157225	0.018996	0.219296	0.1168	0.212074	0.239904	0.212343	0.734458	0.136103	0.192291	0.110247	0.105997	0.281218	0.195551	0.104149	0.041012	0.174013	0.2966479	0.234935	0.208883	0.055861	0.204321			
abcB8	6.664505	7.71405	3.298921	7.429562	11.30949	8.885061	9.780714	6.916769	6.525109	11.07551	11.0921	8.146399	5.2727253	7.354666	6.259928	7.008684	8.160049	7.115313	11.42079	7.129343	9.811637	3.712953	9.161328			
abcB9	6.842912	7.795226	4.36316	8.057427	8.5333891	9.765656	10.25564	3.976807	9.473696	7.5665	5.2727494	6.691976	7.474736	6.527339	7.833424	6.0327394	5.2307284	11.33989	5.270101	8.599939	3.976051	7.62494				
abcB10	2.825061	3.106933	1.136532	2.301583	4.752371	3.201264	3.55259	4.640379	2.339483	1.633918	1.42617	2.380918	3.384194	1.978333	2.546677	3.9108612	1.864886	4.447705	1.538376	1.513867						
abcB11	4.92923	5.926045	2.530428	6.016898	8.145644	7.257995	6.36416	3.383712	4.245162	10.16901	8.433979	4.948483	1.954381	5.200344	6.509304	4.331175	1.016901	5.457363	7.892148	4.303962	6.55196	2.457971	5.433564			
abcC1	6.329446	8.127404	3.298729	7.433735	10.77491	8.893647	8.964642	4.780864	5.50021	13.61978	12.0042	6.8036182	4.932777	6.808653	8.57131	5.022411	1.160266	7.348775	9.434584	6.013751	8.637754	3.402263	9.586915			
abcC2	0.346905	0.577156	0.270287	0.384025	0.4448287	0.650925	0.4549208	0.268497	1.132171	0.572141	0.105614	0.350252	0.256199	0.383886	0.3266451	0.386984	0.388017	0.422901	0.510708	0.291655	0.18947	0.35565				
abcC3	6.013665	7.439689	5.945232	6.209607	8.332764	9.269004	9.845321	4.872408	9.18837	8.198381	7.655851	5.106647	5.919573	6.765684	5.417606	1.678748	6.1487748	9.15554	5.596835	7.878307	3.581028	6.422337				
bulbulin	2.298514	2.087163	1.772524	3.249229	2.136879	2.398394	1.276152	1.406254	3.344242	2.057926	1.278492	1.659775	2.06705	2.043376	1.004109	1.982293	2.084784	2.470466	2.11105	1.445386	2.455287					
abcC4	3.220286	3.368696	1.862558	2.59252	4.503437	3.730238	3.052793	2.10763	3.121299	4.304639	5.198264	3.152392	2.042043	4.142815	2.622131	3.34671	2.05793	2.277096	3.143111	2.225297	4.554299	8.549991	6.111629	8.626731	6.330593	8.475225
abcC5	7.291887	8.497259	7.300674	8.495691	8.914714	10.28172	9.36289	4.897317	6.726237	12.91419	9.846679	7.445036	5.63811	7.616251	8.361338	7.356883	4.697307	8.860292	10.0361	8.428701	10.13339	6.205289	9.101034			
abcC6	5.308255	6.250884	5.862711	5.894473	6.01132	7.391434	6.363759	3.901863	4.50616	8.166663	7.07777	5.604539	4.04964	5.496995	5.453474	5.151823	2.60295	4.688247	6.461338	4.944702	6.983959	4.43756	6.433565			
abcC7	6.225003	6.015751	6.515959	7.435529	7.629163	8.378556	9.326473	3.63326	9.326473	8.788647	8.178847	7.166226	4.766767	6.51443	6.271785	6.575034	3.144826	6.454929	8.549991	6.111629	8.626731	6.330593	8.475225			
abcC8	0.20927	0.312692	0.104774	0.399283	0.316207	0.492878	0.391981	0.090541	0.810137	0.344879	0.5192981	0.254012	0.16607	0.493377	0.347002	0.266	0.196555	0.365982	0.329052	0.330411	0.409944	0.03408	0.161494			
abcC9	4.454158	5.215398	3.603154	4.558374	6.240997	6.4330868	5.816628	3.2492583	4.003031	7.083351	6.895326	5.163141	3.255948	4.606734	4.810717	1.717879	4.781281	6.920567	5.053803	5.490752	3.654303	5.490752				
abcC10	4.421718	4.946461	4.121841	4.067659	5.833341	5.416718	5.4610851	3.4449476	4.77319	6.005754	6.226844	4.169301	2.883633	4.755964	4.331723	4.556479	1.645101	4.42								

abcC1	5.377921	6.014355	4.830902	6.52645	7.497958	7.7118198	7.957887	3.78826	3.902178	8.896234	8.125819	6.0088471	4.413025	5.404135	6.297336	5.365717	1.602617	5.326979	7.10816	5.700667	7.456445	4.850691	7.009039
abcC2	4.550023	5.097199	2.083206	5.287654	6.740772	5.947075	5.545201	2.982741	3.771503	8.8863798	7.793826	4.225222	3.44731	5.073856	5.931241	3.568024	1.343014	4.877474	5.326324	4.41935	6.384966	4.666285	5.233725
abcC3	3.289754	3.800043	1.178961	3.625486	6.3444843	4.852034	4.386753	2.606052	3.18993	5.5714607	7.17708	2.957792	2.417687	3.972703	3.082481	1.772248	0.89427	4.023579	3.746475	3.086382	4.387412	3.645549	4.041307
abcD1	3.324885	3.8888023	1.527821	4.02903	4.8963761	5.135681	5.520253	2.482413	3.418455	5.589764	5.104864	3.723446	2.563685	4.31724	3.968746	3.128986	1.575417	4.202042	5.104484	3.949438	5.473419	3.450191	5.11301
abcD2	2.691791	2.933712	0.973682	2.255953	5.322177	3.067491	3.760776	1.751205	3.294883	3.966396	5.309153	2.865471	1.680424	4.019171	2.339185	3.28914	1.101583	2.982665	3.360074	2.355537	4.726608	1.887609	3.95424
abcD3	2.140153	2.205829	0.720351	1.921719	3.083791	2.384265	2.3388681	1.419681	2.747711	2.203203	3.483447	2.123141	1.126501	3.274346	1.543506	2.419758	1.485437	2.021632	2.86093	1.343676	3.745222	1.837798	2.626086
abcD4	4.14984	3.885758	1.643377	4.797625	5.881954	5.275699	5.142964	2.056191	4.663416	3.004363	6.999542	4.275728	2.8575725	5.542423	4.592518	4.056683	2.764205	4.550051	6.468077	3.353675	6.30511	3.196361	4.226271
abcE1	3.861133	4.032411	1.123749	3.908748	5.426583	4.427617	4.13424	2.162943	3.777744	3.46213	6.370534	3.811598	2.286778	4.590051	3.491907	3.120769	1.952447	3.538173	4.758896	2.553818	5.231677	3.201301	3.925311
abcF1	3.006927	3.174756	1.434718	2.583275	4.162581	3.433896	3.736274	2.009307	3.975386	3.141324	4.572986	2.925539	1.656583	4.641542	2.397631	1.802607	3.19475	4.550298	2.413411	5.138317	2.691841	4.668573	
abcF2	4.790131	5.441651	3.842271	4.674228	5.87547	6.234663	5.246463	3.260485	4.377511	3.836889	6.385178	4.966839	3.338441	5.314146	4.688036	4.673684	2.540565	3.955334	6.320643	4.08181	6.460594	3.856447	6.077975
abcF3	4.204277	4.031992	3.819556	4.154403	6.220227	4.68503	4.375361	3.108019	4.057537	4.083251	6.796562	4.424878	2.649993	5.247922	4.126422	4.173554	2.113369	2.789391	5.566595	3.58488	5.853848	3.048415	4.69793
abcG1	4.681105	5.047226	4.150557	4.956645	6.502208	6.121283	5.5944559	3.498153	4.096599	4.883075	6.594222	4.918009	3.110898	5.169477	4.696716	4.616634	2.266889	1.886996	6.717191	4.143091	6.73207	3.486553	6.323139
abcG2	3.651149	3.601846	1.405222	3.235056	6.296545	4.418258	3.541676	2.341017	4.123002	3.674573	6.905581	4.337574	1.90757	5.424299	3.38139	4.576475	1.676355	1.62062	5.419817	2.920977	5.7438	2.41731	3.91884
abcG3	5.790807	7.229536	3.666519	7.763176	9.081905	8.30557	8.253085	3.405108	4.574626	7.433952	9.335062	7.730029	4.750124	6.333057	7.198438	4.975459	1.482581	1.996986	8.376657	5.91003	7.746507	5.923334	8.29024
abcG5	1.849521	2.2966887	0.447623	1.853678	4.376194	2.754429	2.842199	1.776374	4.73147	3.294423	4.270417	2.125876	0.96839	2.808186	1.864413	1.149096	0.533571	0.697843	2.381646	1.480494	2.896687	1.586751	3.561919
abcG8	3.422902	4.417621	1.874501	4.016039	5.100572	4.995688	4.445349	2.438924	4.338727	4.759615	6.102491	3.924575	1.689189	5.148777	3.708877	3.627643	1.124542	0.889562	6.17538	3.63882	6.247417	3.856546	5.077863

Table 3

	b20	caco2	col6320	lmbt61	lek	lek2	lrepG2	hf75	hf177	lcap	mct7	mda453	mda468	mf29c	stmas	stmbc	stnd2	stnmc	tf7d	tf7f5	mdck				
wt1	0.625575	0.633468	0.668384	0.400289	0.621656	0.621123	0.400733	0.5622468	0.549039	0.390962	0.73678	0.621694	0.507536	0.664708	0.493364	0.844846	0.482902	0.513761	0.39518	0.572626	0.678053	1.246760	0.495741		
abcA1	1.067935	1.31036	0.383940	0.817174	2.135605	1.12835	0.703917	0.846039	1.357568	1.073567	2.459855	1.179514	0.737923	1.595116	1.217115	1.559121	1.197071	0.972456	0.82417	0.82949	1.679435	2.050189	0.862935		
abcA2	1.643533	2.059501	1.365666	1.437249	3.180057	1.928807	1.454453	1.52175	1.832723	1.627496	3.255641	1.922165	1.507406	2.287951	1.9466622	1.844208	1.065716	1.510258	1.024553	1.670174	1.256401	1.014068	1.404345		
sh1	1.390365	1.643217	0.75021	1.34112	2.556266	1.392408	1.504573	1.451991	1.781544	1.285823	3.411175	1.508067	0.998669	2.246812	1.440085	1.534926	0.834975	1.318588	1.121269	1.260814	2.167086	1.885898	1.746449		
abcA3	2.704209	3.345796	2.026091	2.517446	4.345434	2.68124	1.971193	2.397423	2.877423	3.157781	5.656632	2.978834	2.228411	3.1026	3.096152	1.2831308	2.308296	2.726028	2.267062	2.390383	3.581735	4.297679	1.804813		
abcA4	2.103801	2.904362	2.205864	2.217384	3.053411	2.802692	2.23412	3.255694	2.203355	1.99188	3.969112	2.494561	2.089836	2.757477	2.951456	2.803733	0.954946	2.181287	1.907051	2.509148	2.994617	4.311513	1.845699		
abcA5	1.051179	1.349962	0.420592	0.930578	1.193596	1.143116	1.355801	1.121593	1.975003	0.90462	2.669371	1.163195	0.743168	2.1113783	1.132422	1.354663	0.948017	1.013505	0.917291	1.938029	1.860611	1.1119809			
abcA6	1.819137	2.346171	1.041058	2.236483	3.032463	2.4446627	2.108193	1.8338403	2.201172	1.800395	4.192779	2.129047	1.90675	2.599178	2.238761	1.344593	1.142466	1.950437	1.6928	1.884248	2.847344	2.985344	1.62016		
abcA7	0.288957	0.568339	0.198818	0.560872	0.375937	0.442283	0.330229	0.581444	1.543734	0.313801	0.520832	0.293249	0.180035	0.803524	0.373224	0.46228	0.901964	0.268814	0.292069	0.271875	0.388463	0.504119	0.262611		
abcA8	1.338701	1.726724	0.650005	1.601538	2.329269	1.562192	1.104682	1.602261	1.846439	1.28337	3.154813	1.635336	1.128543	2.144792	1.400368	1.592474	1.121732	1.351872	1.074935	1.06233	2.215794	2.331124	1.170965		
abcA9	2.397469	3.075158	1.395623	3.639867	4.616888	3.301872	3.405173	3.52273	2.892316	3.022016	6.372016	2.437209	2.919776	2.719472	3.452203	3.425497	2.043546	1.131171	3.311643	2.821589	2.184486	2.702407	3.661164	2.395371	
abcA10	1.326343	1.927215	0.457038	1.740167	3.017634	1.461084	1.871857	1.520451	1.631431	1.944597	4.042618	1.34173	1.218646	2.130476	1.94012	0.965322	0.575261	1.675096	1.170856	1.320716	2.267016	2.5201	1.861027		
abcA12	1.655644	2.367261	1.131534	2.03472	2.788148	2.042444	1.824851	1.622405	1.815913	1.893298	3.624838	1.972211	1.480277	2.266634	2.256505	1.714529	0.827054	1.755425	1.443137	1.755568	2.4223402	2.282793	2.082827		
abcB1	2.214094	2.709986	1.494853	2.370284	3.412421	2.431277	2.195887	1.562287	1.26583	2.227149	4.615653	2.404548	2.024247	2.790657	2.429782	1.305983	2.212158	1.595806	2.171713	3.036508	0.401019	2.214427			
abcB2	1.667629	2.221349	1.380709	1.664514	2.870063	1.58971	1.739887	1.685054	1.994483	1.821638	3.160028	1.915195	1.425216	2.46317	1.681239	1.889844	1.951877	1.44069	1.910197	1.390459	1.546273	4.486311	1.771761		
abcB3	2.166756	2.501265	1.973617	2.155505	3.2329066	2.615736	1.872813	2.612733	1.844933	2.346864	1.979457	3.838452	2.40551	1.793421	3.015524	1.922764	2.620675	1.650827	2.152817	2.268781	1.558638	3.060383	3.264489		
abcB4	2.730797	3.283257	3.312908	3.677934	4.154998	3.782563	3.835015	3.383875	3.106654	3.091775	5.398659	3.12821	2.731252	3.78884	3.125551	2.8889	2.411262	3.451834	3.519634	2.820545	3.844128	0.052402	2.535107		
abcB6	1.451845	1.95126	1.335779	1.487807	2.267046	1.79348	1.872166	1.563362	1.955536	1.615657	2.864576	1.649393	1.2017	2.223929	1.610864	1.116158	1.215756	1.504506	1.66158	1.312776	2.266957	1.704987	1.849722		
abcB7	0.026344	0.061552	0.008915	0.015592	0.047381	0.086623	0.075751	0.114822	0.342761	0.033199	0.093714	0.041397	0.04744	0.156955	0.076755	0.040478	0.031637	0.068942	0.072051	0.092935	0.076529	0.044888	0.071851		
abcB8	2.368635	3.019983	4.542486	2.561014	4.666834	2.777406	3.088332	3.740167	3.045175	2.628425	5.405767	3.058923	2.284313	3.584394	2.456433	2.723963	1.435114	2.819102	2.775552	2.02905	3.594719	2.983394	3.2211658		
abcB9	2.432043	3.051763	2.282388	2.777405	3.461854	3.052674	3.238294	2.150415	2.461747	2.757258	4.617035	2.841175	2.282323	3.259759	3.040258	2.536886	2.183731	3.363433	1.951987	0.895037	3.150786	3.195011	2.681374		
abcB10	1.004056	1.216337	0.53340	0.703359	1.927944	1.000692	1.028019	1.232321	1.540976	0.866558	2.407709	1.013758	0.708124	2.017916	0.934268	1.315285	0.751222	1.008967	0.949885	0.737711	1.629519	1.236184	1.235681		
abcB11	1.751936	2.320003	1.187586	2.074063	3.304822	2.688797	2.009531	1.829737	1.981156	1.410326	1.838125	1.711692	2.533148	2.554289	3.683335	0.784444	2.162151	1.9117978	1.702549	2.404061	1.975136	1.1910758			
abcC1	2.260214	3.181808	1.548167	2.569312	4.370943	2.780085	2.830645	2.585199	2.566889	3.322217	5.850286	2.55568	2.135203	3.317061	3.363433	1.951987	0.808495	0.811112	0.794946	0.774576	0.783364	2.378907	3.161644	3.457143	3.371319
abcC2	0.123294	0.225951	0.126852	0.132374	0.181832	0.203474	0.144998	0.152025	0.621705	0.13956	0.247874	0.131517	0.110898	0.18797	0.125087	0.150404	0.299319	0.167549	0.124114	0.115372	0.10115	0.095581	0.132		
abcC3	2.13732	2.912573	2.790229	2.14046	3.389264	2.897424	2.16136	2.649476	2.273883	2.241281	3.966213	2.53603	2.210465	2.883492	2.655012	2.105582	1.294998	2.436068	2.16669	2.156999	2.877585	2.258469			
abcC4	0.816916	0.817060	0.831885	0.809782	0.8696845	0.918795	0.757310	0.6900651	0.6562678	0.8838925	1.003349	0.772759	0.737886	0.808495	0.811112	0.794946	0.774576	0.783364	0.506652	0.977261	3.161646	3.863422			
tubulin	1.144524	1.318815	0.87414	0.893897	1.8266862	1.166046	0.963942	1.139678	1.4466651	1.4666651	2.53339	1.258803	0.883919	2.01803	1.028919	1.587505	0.902162	0.76385	0.883164	1.672201	2.341406	1.209116			
abcC5	2.591613	3.326603	3.426568	2.958476	3.616338	3.213392	2.956401	2.64817	3.139039	3.1501054	2.480483	2.10989	2.10308	2.138415	3.494937	2.104478	1.752928	3.281039	3.523533	3.622353	3.510354	2.448125	3.3342	3.675962	4.3200455
abcC6	1.88661	2.447166	2.7515	2.031832	2.428549	2.466806	2.009403	2.10989	2.10308	2.138415	2.493372	2.104478	1.752928	2.67765	2.139967	2.02284	2.07934	1.857434	1.570257	1.956011	2.558662	2.262417			
abcC7	2.21243	2.355114	3.058084	2.562967	3.094842	2.619076	3.008053	3.196573	2.48992	2.310668	4.283174	2.690874	3.174254	2.461086	2.063344	3.174254	2.425294	1.247784	1.907052	1.737432	1.61460	2.308084			
abcC8	0.074577	0.124951	0.049731	0.137633	0.128273	0.15407	0.123771	0.0843959	0.3778079	0.0843959	0.123771	0.104478	1.752928	1.020461	1.975136	1.1910758	0.02786	0.02786	0.02786	0.02786	0.02786	0.02786			
abcC9	1.650946	2.041783	1.69104	1.571217	2.531719	2.01243	1.836706	1.872333	1.868157	1.7283	3.360475	1.938781	1.867111	1.869711	1.867111	1.869711	1.867111	1.867111	1.867111	1.867111	1.867111	1.867111			
abcC10	1.571525	1.956496	1.934471	1.714022	2.176349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349	1.726349				
abcC11	1.91137	2.554568	2.267349	2.249676	3.041617	2.142165	2.512764	1.874144	1.893897	1.8266862	1.166046	0.963942	1.139678	1.241281	2.278674	1.91023	2.263247								

abcC12	1.546044	1.995509	0.977695	1.822661	2.734458	1.859013	1.750938	1.612884	1.760107	2.162109	3.798345	1.586345	1.487875	2.471534	2.327494	1.386732	1.03601	1.932404	1.415934	1.748193	2.339279	3.749558	1.840483	
abcC13	1.169213	1.487684	0.553312	1.24971	2.573845	1.516711	1.353149	1.457866	1.488695	1.457361	1.349772	1.110633	1.046521	1.935151	1.209584	0.669361	0.620419	1.3941	0.910482	1.220902	1.607429	2.927824	1.42116	
abcD1	1.181699	1.522127	0.71704	1.388812	1.985261	1.605577	1.74306	1.342337	1.595344	1.461058	2.487871	1.398128	1.109717	2.102977	1.557359	1.215288	1.664805	1.240521	1.562307	2.005513	1.2.772448	1.798033		
abcD2	0.956691	1.156353	0.45697	0.77763	2.158991	0.958876	1.187492	0.946945	1.537675	0.967506	2.587432	1.075967	0.727389	1.957784	0.91791	1.278342	0.849769	1.181542	0.816577	0.931717	1.731701	1.516814	1.395452	
abcD3	0.760633	0.863563	0.338077	0.662419	1.250969	0.807823	0.738515	0.767677	1.282318	0.537418	1.697669	0.797227	0.487618	1.594972	0.605799	0.940452	1.145877	0.800949	0.695273	0.531527	1.372148	1.476788	0.923485	
edh	0.355411	0.391491	0.469322	0.344701	0.405659	0.312593	0.315757	0.540739	0.466686	0.243926	0.487353	0.375494	0.43286	0.487112	0.392406	0.388655	0.771407	0.396189	0.243024	0.395577	0.366373	0.803564	0.351658	
abcD4	1.474897	1.521249	0.771273	1.653748	2.33607	1.649083	1.623929	1.111863	2.176349	0.732842	3.411248	1.60551	1.236788	2.699782	1.802131	1.576652	2.132327	1.802682	1.571895	1.524425	2.310022	2.56848	1.486204	
abcE1	1.372287	1.578654	0.5274	1.34775	2.201345	1.384041	1.305416	1.160587	1.763019	0.844503	3.104699	1.431232	0.989835	2.233867	1.370245	1.212904	1.506131	1.401787	1.156524	2.049789	1.916738	2.572449	1.380368	
abcF1	1.058693	1.242889	0.673345	0.891148	1.68859	1.07488	1.179796	1.085511	1.856656	0.766625	2.228659	0.98443	0.717069	2.260949	0.940845	1.262637	1.390543	1.265726	1.10583	0.95459	1.882541	2.163066	1.641743	
abcF2	1.702463	2.130359	1.803263	1.611212	2.33344	1.94688	1.655669	1.763072	2.042922	0.935916	3.209307	1.865025	1.445079	2.558852	1.839613	1.816453	1.959809	1.566269	1.536065	1.614467	2.366987	3.098901	1.235612	
abcF3	1.494244	1.57849	1.792602	1.432028	2.523294	1.464507	1.38171	1.686027	1.893594	0.99601	3.312228	1.661515	1.147077	2.556323	1.619232	1.622074	1.630267	1.105128	1.52814	1.418096	2.144692	2.449596	1.677338	
abcG1	1.663714	1.975945	1.947948	1.708562	2.637682	1.913751	1.892763	1.891588	1.911824	1.191108	3.213714	1.846683	1.346584	2.518112	1.843019	1.794228	1.748694	0.747608	1.632436	1.638912	2.466649	2.801668	2.223585	
abcG2	1.297657	1.410091	0.659302	1.115132	2.55255	1.381116	1.11831	1.265879	1.924146	0.896323	3.365456	1.628733	0.832981	2.642239	1.326877	1.778672	1.293175	0.642073	1.317143	1.155471	2.104373	1.942463	1.38268	
abcG4	2.058114	2.8303	1.720778	2.675977	3.685378	2.59632	2.605972	1.841274	1.349131	1.813821	4.793148	2.902579	2.056143	3.085885	2.82471	1.933739	1.143673	0.791185	2.035725	2.337872	2.838111	4.759776	2.91807	
abcG5	0.657339	1.16151	0.210079	0.638965	1.77244	0.861015	0.897445	0.966555	2.208109	0.803595	2.081201	0.798254	0.419177	1.3679	0.731607	0.446602	0.411601	0.276478	0.578796	0.58565	1.061195	1.275059	1.233378	
abcG8	1.216642	1.72946	0.879745	1.384334	2.069095	1.561616	1.403869	1.318827	2.024822	1.160993	2.974068	1.473654	0.735512	2.508029	1.409903	1.454601	1.409903	0.86748	0.352435	1.500072	1.439434	2.288883	3.098981	1.785373

Table 4

abcC12	1.111969	1.214391	1.386991	1.069708	1.335106	1.163744	1.110809	0.987967	1.661499	1.113501	1.052039	1.489838	1.100018	1.616219	0.903452	1.240795	1.463551	1.168004	1.386559	0.079458	1.987208	1.053818	
abcC13	0.840939	0.905348	0.737543	0.950992	1.006877	1.089272	1.020626	1.004046	0.83562	1.133407	1.025338	0.736461	1.047915	0.861287	0.833993	0.456087	0.743055	1.208944	0.751056	0.966344	0.541747	1.55166	0.813723
abcD1	0.849920	0.926309	0.955786	1.056844	0.777017	1.15295	1.158508	0.92448	0.895484	1.136283	0.729333	0.927099	1.111196	0.935983	1.081435	0.792278	1.455509	1.262566	1.023296	1.239125	0.92535	1.469316	1.029512
abcD2	0.688086	0.703713	0.609123	0.591753	0.844588	0.688646	0.789255	0.659217	0.863113	0.755442	0.758516	0.713474	0.728338	0.871361	0.6374	0.832836	1.01774	0.896066	0.673594	0.73898	0.799092	0.803867	0.796192
abcD3	0.547074	0.525532	0.450643	0.504081	0.489974	0.580162	0.490847	0.528706	0.719779	0.417957	0.497679	0.528641	0.488268	0.709882	0.420669	0.612702	1.372378	0.60743	0.573531	0.421575	0.633177	0.782654	0.528766
gdb	0.255624	0.238247	0.625588	0.262307	0.158692	0.224498	0.209865	0.372412	0.261956	0.189704	0.14287	0.24899	0.433437	0.216801	0.272488	0.253208	0.923888	0.300465	0.20047	0.313747	0.169063	0.425855	0.201351
abcD4	1.060798	0.925775	1.028076	1.258453	0.93342	1.184339	1.079328	0.765751	1.221608	0.569994	1.000022	1.064614	1.238456	1.201605	1.251406	1.027184	2.553816	1.367131	1.296636	1.240908	1.065958	1.361219	0.859666
abcE1	0.986998	0.560709	0.703003	1.023294	0.861156	0.993991	0.867632	0.805506	0.989601	0.65678	0.910156	0.94905	0.991174	0.995129	0.951503	0.790204	1.803843	1.063097	0.954016	0.832628	0.884477	1.363322	0.790367
abcF1	0.768642	0.756375	0.897542	0.678137	0.660569	0.771958	0.784113	0.74829	1.042161	0.595922	0.653341	0.728378	0.718025	1.006292	0.653326	0.822604	1.665407	0.959911	0.912198	0.757201	0.868697	1.146361	0.940024
abcF2	1.224471	1.296456	2.403678	1.2266085	0.932291	1.399647	1.101049	1.214244	1.146714	0.7277873	0.940822	1.236699	1.447004	1.152113	1.277433	1.183414	2.347198	1.187838	1.2671	1.280657	1.092244	1.642326	1.222802
abcF3	1.074713	1.060609	2.389467	1.089731	0.987102	1.05178	0.91834	1.157463	1.062395	0.774609	0.970993	1.101751	1.148605	1.137756	1.1244	1.056777	1.952517	0.833814	1.115936	1.124746	0.989666	1.298214	0.960405
abcG1	1.196602	1.202485	2.596537	1.300165	1.031185	1.374418	1.258006	1.302754	1.073128	0.926339	0.942114	1.224336	1.348379	1.120749	1.279799	1.168968	2.094352	0.566976	1.346596	1.299883	1.138141	1.484801	1.273173
abcG2	0.933321	0.858128	0.87909	0.848582	0.999213	0.99189	0.743274	0.871823	1.080044	0.697082	0.985597	1.080013	0.854118	1.175995	0.921188	1.1588	1.548793	0.48694	1.086511	0.916448	0.971061	1.029448	0.791691
abcG4	1.480268	1.722414	2.293729	2.036339	1.441704	1.864626	1.732034	1.268103	1.198349	1.410631	1.405131	1.924701	2.058888	1.373451	1.961488	1.259826	1.369739	0.600024	1.679269	1.854256	1.309644	2.522541	1.670819
abcG5	0.472781	0.706831	0.280027	0.486234	0.694468	0.618364	0.5964478	0.661543	1.239435	0.624966	0.610113	0.529322	0.419736	0.608818	0.50803	0.29096	0.49296	0.209677	0.477449	0.464501	0.489687	0.675744	0.717655
abcG8	0.875052	1.052484	1.172665	1.053437	0.809421	1.121522	0.933028	0.908288	1.136554	0.902918	0.87186	0.97718	0.736492	1.116261	1.010079	0.918548	1.03951	0.267282	1.237409	1.14167	1.056204	1.642369	1.022435

Table 5

	dox 0h	dox 2h	dox 4h	dox 8h
abcA1	1.793411	3.052731	1.865644	2.34586
abcA2	3.394744	6.223801	2.94659	4.02209
abcA3	4.445693	8.071446	4.290698	5.179128
abcA4	5.098287	8.764862	4.534571	6.09907
abcA5	2.006987	3.30202	2.020768	2.451236
abcA6	3.567858	6.044507	3.366697	4.295772
abcA7	0.906336	1.841564	0.932998	0.935086
abcA8	1.575163	3.035544	1.785517	2.17441
abcA9	5.12988	7.825115	4.816535	5.72013
abcA10	3.225933	4.820089	3.418986	3.792907
abcA12	3.485887	5.828746	3.418674	4.195394
abcB1	3.658465	6.734501	3.865342	4.758501
abcB2	2.792672	5.067235	3.714749	4.008349
abcB3	3.312315	6.838271	4.325461	4.812997
abcB4	5.149497	9.148426	5.624165	6.417042
abcB6	2.795918	5.173665	3.283246	3.61177
abcB7	0.143706	0.262199	0.161948	0.176608
abcB8	4.688411	8.003626	5.437681	5.35345
abcB9	4.532227	8.387881	4.897126	5.522502
abcB10	1.264095	2.115507	1.484423	1.542031
abcB11	3.285622	5.310097	3.404054	3.888131
abcC1	4.397451	7.004924	4.767338	5.055766
abcC2	0.340701	0.614144	0.360272	0.354806
abcC3	4.024623	7.155717	4.013536	4.199702
abcC4	1.480616	2.612061	2.089878	2.285607
abcC5	5.251928	10.50642	6.290367	6.521707
abcC6	3.94515	7.696336	4.515506	4.899487
abcC7	3.904822	7.480766	4.5794	5.093014
abcC8	0.210057	0.322281	0.243749	0.22546
abcC9	3.239867	5.598434	3.67832	3.981505
abcC10	3.504958	5.15091	3.334564	3.632591
abcC11	4.300962	7.608052	4.383947	5.056108
abcC12	2.421183	5.226012	3.53205	3.976487
abcC13	2.231485	3.20307	2.54815	2.593022
abcD1	2.923938	4.476831	3.385873	3.516307
abcD2	1.810003	2.503156	2.516228	2.409319
abcD3	1.143253	2.09855	1.78719	1.733079
abcD4	2.411452	4.360857	3.102722	3.194107
abcE1	2.060757	4.155317	2.79372	3.087661
abcF1	1.969904	2.485367	2.869902	2.661525
abcF2	3.671255	5.978677	4.068172	4.806913
abcF3	2.398669	3.920654	2.794743	2.793001
abcG1	3.224847	5.471919	3.555021	3.838933
abcG2	1.711538	2.988958	2.136826	2.080252
abcG4	5.107502	9.589581	5.308586	6.270866
abcG5	1.427298	2.200836	1.76435	1.751627
abcG8	2.379986	4.696989	2.811059	2.9413
standard	11	11	11	11

Table 6

	vin0h	vin2h	vin4h	vin 8h
abcA1	6.033981	6.833133	5.063992	6.364167
abcA2	6.092914	7.398232	6.087334	9.54274
abcA3	8.389483	10.83098	9.241369	14.93551
abcA4	8.853516	11.00906	9.361	15.90913
abcA5	7.368576	7.894724	6.136832	8.641923
abcA6	8.249337	9.900094	7.850423	14.54007
abcA7	3.030993	2.265104	1.807985	3.13492
abcA8	6.552532	8.424365	6.723552	10.25917
abcA9	10.06712	12.25496	10.20161	18.62193
abcA10	7.746441	10.27696	8.658689	10.49502
abcA12	6.787256	8.473897	6.774483	8.088676
abcB1	9.188582	12.11622	9.658148	11.52253
abcB2	6.87262	8.667108	8.938251	8.490746
abcB3	8.053858	9.144091	8.399529	14.78834
abcB4	11.54316	16.19463	13.24367	23.08427
abcB6	6.745944	8.654504	8.468344	7.559171
abcB7	0.969163	0.461908	0.346696	0.695811
abcB8	12.78564	16.52333	15.3408	20.22378
abcB9	9.119206	12.61687	12.27524	12.23283
abcB10	4.908251	4.258718	3.989921	5.497747
abcB11	5.160003	7.105947	7.457957	9.052987
abcC1	11.60798	16.22045	15.1016	12.68866
abcC2	2.150775	1.429625	1.196853	1.419261
abcC3	9.195576	12.14918	10.16914	11.28798
abcC4	5.334611	5.277331	5.742927	6.273209
abcC5	9.43692	10.94563	12.64342	17.32233
abcC6	7.241802	8.103238	8.42442	9.917254
abcC7	11.61626	13.77896	13.42656	16.12312
abcC8	0.527266	0.353293	0.453532	0.880544
abcC9	7.904892	9.880234	9.013857	9.735132
abcC10	8.724349	10.91895	8.808216	9.006965
abcC11	8.108411	9.016067	7.820571	11.10364
abcC12	7.448083	8.04745	7.567471	8.523092
abcC13	7.931162	9.550546	8.963819	7.491041
abcD1	8.371099	10.01996	9.667713	8.253821
abcD2	6.641271	6.669278	6.511982	5.888302
abcD3	5.873059	5.918177	5.802474	5.647165
abcD4	7.456156	8.051998	8.68655	10.27714
abcE1	6.417	6.427651	6.814744	7.53135
abcF1	7.793391	7.85194	7.234306	5.758539
abcF2	7.663223	9.20984	8.64578	9.39799
abcF3	8.043509	8.975626	7.819995	8.132801
abcG1	7.649376	9.753252	8.36374	8.726646
abcG2	5.486492	6.441959	5.659027	6.036638
abcG4	9.481624	12.50021	13.2242	9.999301
abcG5	5.764325	6.024204	6.084062	4.098926
abcG8	7.991649	9.698668	8.750869	7.315558
standard	24	24	24	24